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(54) Title: PLASMID VECTORS FOR TRANSFORMATION OF FILAMENTOUS FUNGI

(57) Abstract: The invention relates to novel plasmid vectors for transformation of filamentous fungi and to a method of modifying the genome of filamentous fungi based on these vectors. The invention furthermore relates to the modification of a specific gene via the process of homologous recombination, to recombinant expression of foreign genes in filamentous fungi and to new selection markers for detecting successful transfer of the target gene in filamentous fungi.

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# Plasmid vectors for transformation of filamentous fungi

## Description

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The invention relates to novel plasmid vectors for transformation of filamentous fungi and to a method of modifying the genome of filamentous fungi based on these vectors. The invention furthermore relates to the modification of a specific gene via the process of homologous recombination, to recombinant expression of foreign genes in filamentous fungi and to new selection markers for detecting the successful transfer of the target gene in filamentous fungi.

10 One method currently used for transformation of filamentous fungi is random mutagenesis based on transposons insertion, a method also known for plant transformation (WO 01/38509). This method allows the genomes of several species such as *Magnaporthe grisea* to be studied (for examples WO 00/55346; WO 00/56902). However, 15 this strategy requires a big effort in terms of bioinformatic tools and molecular biology to localise precisely the insertion in the genome.

Alternatively, known transformation methods are based on targeted 25 integration. Targeted transformation of fungi can be carried out either by offering a knock-out cassette with a marker-gene flanked by two homologous sequences (Aronson et al, 1994, Mol. Gen. Genet. 242: 490-494; Royer et al, 1999, Fungal Genetics and Biology 28: 68-78; Schaefer, 2001, Current Opinion in Plant Bio- 30 logy 4: 143-150) or by quoting a plasmid with the marker gene in the neighborhood of a homologous sequence (Shortle et al., 1982, Science 217: 371-373; Bird and Bradshaw, 1997, Mol Gen Genet. 255: 219-225; Feng et al., 2001, Infection and Immunity 69 (3): 1781-1794; Schaefer, 2001, Current Opinion in Plant Biology 4: 35 143-150). Both procedures are in principle attractive methods to study the gene function, but they have the disadvantage of a high frequency of integration at ectopic sites by illegitimate recombination. The gene targeting efficiency (gene targeting / gene targeting + illegitimate recombination) is 95% for *S. cerevisiae*, 40 10-90% for *S. pombe*, 5-75% for *Aspergillus nidulans* and 1-30% for *Neurospora crassa* using a size of homology of 2-9 Kb (Schaefer, 2001, Current Opinion in Plant Biology 4: 143-150). Especially for filamentous fungi this side effect is quite high, if conventional plasmid vectors are used.

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In addition, the efficiency of the gene targeting increases if the length of homologous recombination region is increased (Schaefer, 2001, Current Opinion in Plant Biology 4: 143-150). Thus, plasmid vectors currently used comprise gene fragments of the  
5 gene to be knocked out of a size of at least 2000bp as indicated above. The overall size of these plasmid vectors is at least 8000bp (P. J. Punt et. al., 1992, Methods in Enzymology, vol.216, pp 447-457; ; Schaefer, 2001, Current Opinion in Plant Biology 4: 143-150). Since transformation efficiency decreases with the in-  
10 crease of the plasmid vector size, transformation efficiency is unsatisfactory giving rise to long times until positive clones can be identified. This is an obstacle especially to large scale genomic analysis projects or recombinant expression.

15 Furthermore, currently used plasmid vectors contain many unique restriction sites, causing difficulties in construction of the knock-out (KO-) plasmids and the transformation process. The efficiency of homologous recombination is improved when the KO-plasmid is digested with a restriction enzyme presenting a unique  
20 site in the middle of a DNA fragment homologous to the targeted gene. The presence of high amounts of restriction sites especially unique ones in the plasmid backbone decreases the chance of finding a natural restriction site in the appropriate location of the targeted DNA fragment. This problem is usually overcome by  
25 modification of the targeted DNA fragment requiring several cloning steps and additional manipulation in terms of molecular biology, which is a disadvantageous and time consuming methodology.

Integration of recombinant gen by homologous recombination in  
30 fungi is also a tool to identify gene functions for essential genes: the biochemical characterization of an essential gene cannot be studied by classical knock-out strategy since the mutants carrying a disruption of such a gene are not viable. One way to overexpress such a gene overcomes the problem when a typical phe-  
35 notype can be assigned to the mutant that overexpresses the gene. Another approach can be to regulate the gene expression by an inducible promoter sequence so that the gene could be expressed or repressed when needed and consequently permits viable mutants to be isolated. As mentioned above, these approaches require at  
40 least several thousand bp of the nucleic acid sequence to be studied that need to be integrated in the genome of the fungi together with a plasmid vector comprising the different parts of the nucleic acid sequence. In addition, if the recombinant DNA is integrated at an ectopic site, the identification of the mutant  
45 strains becomes more complicated and the position of the integration in the genome may influence the level of expression of the recombinant protein. Taking the aforesaid into consideration,

currently existing plasmid vectors currently used for transformation of filamentous fungi exhibit a lot of disadvantages and are neither suitable for large scale analysis e.g. in functional genomic studies nor convenient for recombinant expression in a filamentous fungi. Additionally, there is a constant need for new selection markers facilitating the selection process.

Thus, object of the present invention was to develop tools for targeted transformation of filamentous fungi that overcome the disadvantages of the state of the art like plasmid vectors suitable for functional genomic studies and recombinant expression and new selection markers.

We have found that the object of the invention is achieved by construction of a plasmid vector for targeted transformation of filamentous fungi comprising

- a) an origin of replication for a host organism not originating from the filamentous fungi to be transformed;
  - b) a selection marker for a host organism not originating from the filamentous fungi to be transformed;
  - c) a promotor facilitating recombinant expression in filamentous fungi that is functionally linked to the coding region of the hygromycin resistance gene which is functionally linked to a terminator which facilitates transcription termination in filamentous fungi;
- wherein the overall size of the elements a), b) and c) does not exceed 4500 bp; and
- d) a nucleic acid sequence, which is homologous to nucleic acid sequences of the filamentous fungi to be transformed and makes homologous recombination in the filamentous fungi to be transformed possible.

The term overall size of the elements a), b) and c) designates the combination of the essential elements of the expression vector without the nucleic acid sequence d).

The overall size of the elements a), b) and c) does not exceed 4500 bp, preferably 4100 bp, more preferably 3700 bp.

In addition to the nucleic acid elements a), b), c) and d), the plasmid vector optionally comprises a cloning site containing rare restriction sites or a TA-cloning site by which further nu-

cleic acid sequences can be cloned easily into the plasmid vector. A TA-cloning site comprises thymidine residues linked onto the 3'-ends of linearized plasmid DNA, which would allow some annealing to occur between the vector and the A-tailed PCR product to be ligated. This process is called TA cloning. Preferably, the vector is modified in such a way that there are only few unique restriction sites left enabling the digestion by commercially available restriction enzymes of the homologous sequence of the targeted gene prior to the transformation.

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Filamentous fungi that can be transformed with the vectors of the present invention are non-phytopathogenic filamentous fungi e.g. *Neurospora* species like *Neurospora crassa* and phytopathogenic filamentous fungi the phytopathogenic filamentous fungi being preferred. Examples of other non-phytopathogenic filamentous fungi are *Aspergillus* species such as *Aspergillus parasiticus*, *Aspergillus nidulans*, *Aspergillus niger* and *Wangiella* such as *Wangiella dermatidis*. Preferred phytopathogenic filamentous fungi are selected from the group consisting of the genera *Alternaria*, *Podosphaera*, *Sclerotinia*, *Physalospora*, *Botrytis*, *Corynespora*, *Colletotrichum*, *Diplocarpon*, *Elsinoe*, *Diaporthe*, *Sphaerotheca*, *Cinula*, *Cercospora*, *Erysiphe*, *Sphaerotheca*, *Leveillula*, *Mycosphaerella*, *Phyllactinia*, *Gloesporium*, *Gymnosporangium*, *Leptotthrydium*, *Podosphaera*, *Gloedes*, *Cladosporium*, *Phomopsis*, *Phytopora*, *Phytophthora*, *Erysiphe*, *Fusarium*, *Verticillium*, *Glomerella*, *Drechslera*, *Bipolaris*, *Personospora*, *Phaeoisariopsis*, *Spaceloma*, *Pseudocercospora*, *Pseudoperonospora*, *Puccinia*, *Typhula*, *Pyricularia*, *Rhizoctonia*, *Stachosporium*, *Uncinula*, *Ustilago*, *Gaeumannomyces* and *Fusarium*, more preferred from the group consisting of the genera and species *Alternaria*, *Podosphaera*, *Sclerotinia*, *Physalospora* such as *Physalospora* canker, *Botrytis* species such as *Botrytis cinerea*, *Corynespora* such as *Corynespora melonis*, *Colletotrichum*, *Diplocarpon* such as *Diplocarpon rosae*, *Elsinoe* such as *Elsinoe fawcetti*, *Diaporthe* such as *Diaporthe citri*, *Sphaerotheca*, *Cinula* such as *Cinula neccata*, *Cercospora*, *Erysiphe* such as *Erysiphe cichoracearum* and *Erysiphe graminis*, *Sphaerotheca* such as *Sphaerotheca fuliginea*, *Leveillula* such as *Leveillula taurica*, *Mycosphaerella*, *Phyllactinia* such as *Phyllactinia kakiicola*, *Gloesporium* such as *Gloesporium kaki*, *Gymnosporangium* such as *Gymnosporangium yamadae*, *Leptotthrydium* such as *Leptotthrydium pomi*, *Podosphaera* such as *Podosphaera leucotricha*, *Gloedes* such as *Gloedes pomigena*, *Cladosporium* such as *Cladosporium carpophilum*, *Phomopsis*, *Phytopora*, *Phytophthora* such as *Phytophthora infestans*, *Verticillium*, *Glomerella* such as *Glomerella cingulata*, *Drechslera*, *Bipolaris*, *Personospora*, *Phaeoisariopsis* such as *Phaeoisariopsis vitis*, *Spaceloma* such as *Spaceloma ampelina*, *Pseudocercospora* such as *Pseudocercospora herpotrichoides*;

Pseudoperonospora; Puccinia; Typhula; Pyricularia such as Pyricularia oryzae; Rhizoctonia; Stachosporium such as Stachosporium nodorum; Uncinula such as Uncinula necator; Ustilago; Gaeumannomyces species such as Gaeumannomyces graminis and Fusarium such as Fusarium dimerium, Fusarium merismoides, Fusarium lateritium, Fusarium decemcellulare, Fusarium poae, Fusarium tricinctum, Fusarium sporotrichioides, Fusarium chlamydosporum, Fusarium moniliforme, Fusarium proliferatum, Fusarium anthophilum, Fusarium subglutinans, Fusarium nygamai, Fusarium oxysporum, Fusarium solani, Fusarium culmorum, Fusarium sambucinum, Fusarium crookwellense, Fusarium avenaceum ssp. avenaceum, Fusarium avenaceum ssp. aywerte, Fusarium avenaceum ssp. nurragi, Fusarium heterosporum, Fusarium acuminatum ssp. acuminatum, Fusarium acuminatum ssp. armeniacum, Fusarium longipes, Fusarium compactum, Fusarium equiseti, Fusarium scripi, Fusarium polyphialidicum, Fusarium semitectum and Fusarium beomiforme and especially preferred from the genera Fusarium such as Fusarium graminearum, most preferred from the group consisting of the genera and species Fusarium, Fusarium dimerium, Fusarium merismoides, Fusarium lateritium, Fusarium decemcellulare, Fusarium poae, Fusarium tricinctum, Fusarium sporotrichioides, Fusarium chlamydosporum, Fusarium moniliforme, Fusarium proliferatum, Fusarium anthophilum, Fusarium subglutinans, Fusarium nygamai, Fusarium oxysporum, Fusarium solani, Fusarium culmorum, Fusarium sambucinum, Fusarium crookwellense, Fusarium avenaceum ssp. avenaceum, Fusarium avenaceum ssp. aywerte, Fusarium avenaceum ssp. nurragi, Fusarium heterosporum, Fusarium acuminatum ssp. acuminatum, Fusarium acuminatum ssp. armeniacum, Fusarium longipes, Fusarium compactum, Fusarium equiseti, Fusarium scripi, Fusarium polyphialidicum, Fusarium semitectum and Fusarium beomiforme wherein Fusarium graminearum is most preferred.

The host organism in which the origin of replication a) is functionally active essentially serves to construct and propagate the plasmid vector of the invention. The host organism must be genetically different from the filamentous fungi to be transformed, since replication of the plasmid vector should not take place in the filamentous fungi to be transformed but is desired in the host organism, due to the use of the origin of replication a).

Host organisms which may be used are all common microorganisms which can easily be manipulated by genetic engineering. Preferred host organisms are Gram-negative bacteria such as the genera Escherichia and Salmonella e.g. Escherichia coli and Salmonella typhimurium or Gram-positive bacteria such as the genera Bacillus and Streptomyces, e.g. Bacillus subtilis and Streptomyces nidulans. Particularly preferred are gram-negative bacteria such as

Escherichia, e.g. Escherichia coli.

Preferred origins of replication (ori) are the col E1 ori, the fl ori.

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The term "selection marker for a host organism" set forth in b) means a gene or the expression product of the gene. Preferred meanings are genes whose expression causes resistance of the host organism to antibiotics, by preference resistance to kanamycin, chloramphenicol, tetracycline, zeocin or ampicillin, and particularly preferred ampicillin and kanamycin.

In a preferred embodiment, the element a) of the plasmid vector according to the invention comprises a col E1 origin of replication and the ampicillin resistance gene as selection marker for the host organism.

The element c) is hereinbelow termed "hygromycin cassette". The coding region of the hygromycin resistance gene (hereinbelow termed "hygromycin gene") is known by the skilled artisan (Gritz L. and Davies J., 1983, Gene 25, 179-188, Kaster, K.R., Burgett S.G. and Ingolia T.D., 1984, Curr. Genet. 8, 353-358) and has a length of 1026bp.

Examples of suitable promoters to which the coding region of the hygromycin gene is functionally linked, are the GPD-1-, PX6-, TEF-, CUP1-, PGK-, GAP1-, TPI, PHO5-, AOX1, GAL10/CYC-1, CYC1, OliC-, ADH-, TDH-, Kex2-, MF $\alpha$ -, or the NMT-promotor (Degryse et al., Yeast 1995 Jun 15; 11(7):629-40; Romanos et al. Yeast 1992 Jun; 8(6):423-88; Benito et al. Eur. J. Plant Pathol. 104, 207-220 (1998); Cregg et al. Biotechnology (N Y) 1993 Aug; 11(8):905-10; Luo X., Gene 1995 Sep 22; 163(1):127-31; Nacken et al., Gene 1996 Oct 10; 175(1-2):253-60; Turgeon et al., Mol Cell Biol 1987 Sep; 7(9):3297-305), preferably the CYC1-, ADH-, TDH-, Kex2-, GPD-1-, PX6, TEF-, CUP1-, PGK-, GAP1-, TPI, PHO5- or AOX1-promotor, more preferably the GPD-1-, PX6, TEF- or the CUP1-promotor, most preferably the GPD1 or the TEF-promotor.

Examples of suitable terminators that are functionally linked to the coding region of the hygromycin gene are the AOX1-, nos-, PGK-, TrpC- or the CYC1-terminator (Degryse et al., Yeast 1995 Jun 15; 11(7):629-40; Brunelli et al. Yeast 1993 Dec 9(12): 1309-18; Frisch et al., Plant Mol. Biol. 27 (2), 405-409 (1995); Scorer et al., Biotechnology (N.Y.) 12 (2), 181-184 (1994), Genbank acc. number Z46232; Punt et al., (1987) Gene 56 (1),



117-124)), preferably the CYC1- or nos-terminator, more preferably the nos-terminator.

In a preferred embodiment, the hygromycin cassette comprises a  
5 GPD-1 promotor functionally linked to the coding region of the hygromycin region which is functionally linked to the nos-terminator.

A functional linkage is understood to mean the sequential arrangement of promoter and coding sequence, of coding sequence and terminator or of promoter, coding sequence and terminator in such a manner that each of the regulatory elements can, upon expression of the coding sequence, fulfil its function for the recombinant expression of the nucleic acid sequence. Direct linkage in  
10 the chemical sense is not necessarily required for this purpose. Preferred arrangements are those in which the hygromycin gene to be expressed recombinantly is positioned downstream of the sequence which acts as promoter, so that the two sequences are linked covalently to each other. The distance between the promoter sequence and the nucleic acid sequence to be expressed recombinantly is preferably less than 100 base pairs, especially preferably less than 50 base pairs, and very especially preferably less than 10 base pairs. The distance between the terminator sequence and the nucleic acid sequence to be expressed recombinantly is preferably less than 100 base pairs, especially preferably less than 50 base pairs, and very especially preferably less than 10 base pairs. However, further sequences which, for example, exert the function of a linker with certain restriction enzyme cleavage sites, or of a signal peptide, may also be positioned between the two sequences.  
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These vectors are not only much more smaller than the currently used plasmid vectors, but also exhibit a high transformation efficiency. Surprisingly, a high transformation efficiency can be  
35 gained even if small DNA-fragments of at least 300bp, preferably at least 400bp, more preferably at least 450bp, most preferably at least 500bp of the nucleic acid sequence d) to be analyzed are used. The average degree of illegitimate recombination is below 30%, preferably below 25%, more preferably 20% and most preferably between 0 and 15%.  
40

The nucleic acid sequence d) has a homology of at least 80% to the nucleic acid sequence of the filamentous fungi to be transformed, preferably at least 90%, more preferably at least 95% and  
45 most preferably at least 100%.

In a preferred embodiment, the nucleic acid sequence d) originates from a filamentous fungi and has a length of at least 300bp, preferably 400bp, more preferably at least 450bp, and most preferably at least 500bp. These lengths are suitable for functional genomic studies for which a high number of transformants is required. Also nucleic acid sequences exceeding 500bp can be used, e.g. for the purpose of recombinant expression.

If the nucleic acid sequence d) is to be expressed recombinantly in the filamentous fungi, it can be functionally linked to a promoter e) and optionally to a terminator f).

Examples of suitable promoters e) are for example the AUG1-, GPD-1-, PX6-, TEF-, CUP1-, PGK-, GAP1-, TPI, PHO5-, AOX1, GAL10/CYC-1, CYC1, OliC-, ADH-, TDH-, Kex2-, MF $\alpha$ - or the NMT-promotor or combinations of the aforementioned promoters (Degryse et al., Yeast 1995 Jun 15; 11(7):629-40; Romanos et al. Yeast 1992 Jun;8(6):423-88; Benito et al. Eur. J. Plant Pathol. 104, 207-220 (1998); Cregg et al. Biotechnology (N Y) 1993 Aug;11(8):905-10; Luo X., Gene 1995 Sep 22;163(1):127-31; Nacken et al., Gene 1996 Oct 10;175(1-2): 253-60; Turgeon et al., Mol Cell Biol 1987 Sep;7(9):3297-305).

Examples of suitable terminators f) are the NMT-, Gcyl-, TrpC-, AOX1-, nos-, the PGK- or the CYC1-terminator (Degryse et al., Yeast 1995 Jun 15; 11(7):629-40; Brunelli et al. Yeast 1993 Dec9(12): 1309-18; Frisch et al., Plant Mol. Biol. 27 (2), 405-409 (1995); Scorer et al., Biotechnology (N.Y.) 12 (2), 181-184 (1994), Genbank acc. number Z46232; Zhao et al. Genbank acc number : AF049064; Punt et al., (1987) Gene 56 (1), 117-124).

The nucleic acid sequence d) can be also functionally linked to an affinity tag to purify the encoded protein and/or to a reporter gene to study biochemical properties of the nucleic acid sequence d) in vivo, respectively.

"Reporter genes" encode readily quantifiable proteins. Using these genes, an assessment of transformation efficacy or of the site or time of expression can be made via growth, fluorescence, chemoluminescence, bioluminescence or resistance assay or via photometric measurement (intrinsic color) or enzyme activity. Very especially preferred in this context are reporter proteins (Schenborn E, Groskreutz D. Mol. Biotechnol. 1999; 13(1):29-44) such as the "green fluorescence protein" (GFP) (Gerdes HH and Kaether C, FEBS Lett. 1996; 389(1):44-47; Chui WL et al., Curr. Biol. 1996, 6:325-330; Leffell SM et al., Biotechniques.

23(5):912-8, 1997), chloramphenicol acetyl transferase, a luciferase (Giacomin, Plant Sci. 1996, 116:59-72; Scikantha, J. Bact. 1996, 178:121; Millar et al., Plant Mol. Biol. Rep. 1992 10:324-414), and luciferase genes, in general  $\beta$ -galactosidase or  $\beta$ -glucuronidase (Jefferson et al., EMBO J. 1987, 6, 3901-3907), the Ura3 gene, the Ilv2 gene, the 2-desoxyglucose-6-phosphate phosphatase gene,  $\beta$ -lactamase gene, the neomycin phosphotransferase gene, the hygromycin phosphotransferase gene, or the BASTA (= gluphosinate) resistance gene.

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The term "affinity tag" denotes a peptide or polypeptide whose coding nucleic acid sequence can be fused to the nucleic acid sequence d) either directly or using a linker, by classical cloning techniques. The affinity tag serves to isolate the recombinant target protein by means of affinity chromatography. The abovementioned linker can optionally comprise a protease cleavage site (for example for thrombin or factor Xa), whereby the affinity tag can be cleaved off from the target protein, as required. Examples of customary affinity tags are the "his-tag", for example from Quiagen, Hilden, "strep-tag", "myc-tag" (Invitrogen, Carlsberg), New England Biolab's tag which consists of a chitin binding domain and an intein, and what is known as the CBD-tag from Novagen.

25 In a particularly preferred embodiment, the plasmid vector comprises an coli E1 ori, the ampicillin resistance gene as selection marker, a GPD-1 promotor functionally linked to the coding region of the hygromycin resistance gene which is functionally linked to the nos-terminator.

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Preferably, the vector also comprises a multiple cloning site comprising an appropriate restriction enzyme site. Appropriate restriction sites are well known by the skilled artisan.

35 In a further preferred embodiment, the plasmid vector additionally comprises a TA-cloning site to facilitate the overall cloning procedure.

40 Examples of particularly preferred embodiments are set forth in Fig. 1 and 2.

All of the above mentioned embodiments of plasmid vectors are hereinbelow termed "plasmid vector (or vector) according to the invention".

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## 10

A vector according to the invention may also comprise at least one additional selection marker.

If a plasmid is used for recombinant expression in host organisms, a marker is required indicating the successful transfer of the plasmid vector DNA into the filamentous fungi to be transformed.

Surprisingly, we have found that the gene fragments of the polyketide synthase are a suitable selection marker. The term "selection marker" referring to the polyketide synthase herein means a nucleic acid sequence.

More precisely, the term "selectable marker", "selection marker" or "marker" used in connection with polyketide synthetase for transformation of filamentous fungi means a nucleic acid sequence encoding a polyketide synthetase or fragments of the aforementioned nucleic acid sequence. Preferred embodiments of the aforementioned marker as well as preferred embodiments of methods of use of the respective marker are described herein below.

Polyketide synthases are multifunctional enzymes that are involved in the biosynthesis of several important polyketides. Polyketides constitute a large and highly diverse group of secondary metabolites, synthesized by bacteria, fungi and plants and algae. They include antibiotics, compounds with mycotoxic activity, and compounds within pigment biosynthetic pathways. Furthermore a polyketide synthase is described to be required for fungal virulence of *Cochliobolus heterostrophus* toward maize (Yang et al., 1996 PMID:8953776). Polyketide Synthetases are furthermore known from *Wangiella dermatidis* (PubMedID:11179356), from *Aspergillus nidulans* (Swiss-prot ID: Q03149) and from *Aspergillus parasiticus* (Swiss-Prot ID:Q12053).

The use of polyketide synthase as selectable marker for recombinant expression in filamentous fungi has not yet been described.

The present invention also encompasses a selection marker comprising a nucleic acid sequence encoding a polyketide synthetase fragment, wherein said nucleic acid sequence comprises

- i. a nucleic acid sequence shown in SEQ ID NO:1 or SEQ ID NO:2; or
- ii. parts of the nucleic acid sequence as defined in i. consisting of at least 300bp.

## 11

Parts or segments of nucleic acid sequences set forth in ii. consist of at least 300bp, preferably at least 400bp, more preferably at least 450bp, most preferably at least 500bp of the nucleic acid sequences. In a further preferred embodiment, those 5 parts are selected from SEQ ID NO:1, preferably from 732bp to 5881bp of SEQ ID NO:1 e.g. from 2236bp to 2870bp.

Furthermore, the present invention encompasses a selection marker comprising a nucleic acid sequence encoding a polyketide synthetase or a polyketide synthetase fragment, wherein said nucleic acid sequence comprises

- 15 i. a nucleic acid sequence shown in SEQ ID NO:3, SEQ ID NO:4 or SEQ ID NO:5; or
- ii. a nucleic acid sequence which, owing to the degeneracy of the genetic code, can be deduced from the amino acid sequence shown in SEQ ID NO:6 by back translation; or
- 20 iii. a functional equivalent of the nucleic acid sequence set forth in i) which is encoded by an amino acid sequence that has at least an identity of 50% with the SEQ ID NO:6; or
- 25 iv. parts of the nucleic acid sequence as defined in i., ii. or iii. consisting of at least 300bp.
- v. parts of the nucleic acid sequence as defined in i., ii. or iii. consisting of at least 300bp comprising
- 30 a) a nucleic acid sequence shown in SEQ ID NO:7; or
- b) a nucleic acid sequence which, owing to the degeneracy of the genetic code, can be deduced from the amino acid sequence shown in SEQ ID NO:8 by back translation; or
- 35 c) a functional equivalent of a nucleic acid sequence set forth in a) which is encoded by an amino acid sequence that has at least an identity of 85% with the SEQ ID NO:8.

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Parts or segments of nucleic acid sequences set forth in iii. or v. consist of at least 300bp, preferably at least 400bp, more preferably at least 450bp, most preferably at least 500bp of the nucleic acid sequences. Preferably, the aforementioned parts or 45 segments of nucleic acid sequences are those set forth in v.a), v.b) or v.c), more preferably those set forth in v.a) or v.b) and most preferably those set forth in v.a). For example, those parts

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can be selected from 2234bp to 2865bp of SEQ ID NO:3.

The functional equivalents of the nucleic acid sequence set forth in iv. are encoded by an amino acid sequence that has at least an identity of 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64% or 65% or preferably of 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78% or 79% more preferably of 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89% or 90% most preferably of 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98% or 99% with SEQ ID NO:6.

The functional equivalents of the nucleic acid sequence set forth in v.c) are encoded by an amino acid sequence that has at least an identity of 85%, 86%, 87% or 88% or preferably of 89%, 90%, 91%, 92% or 93% more preferably of 94%, 95% or 96% most preferably of 97%, 98% or 99% with SEQ ID NO:8.

Preferred are nucleic acid sequences as defined above originating from filamentous fungi, preferably phytopathogenic filamentous fungi selected from the group consisting of the genera *Neurospora*, *Alternaria*, *Podosphaera*, *Sclerotinia*, *Physalospora*, *Botrytis*, *Corynespora*; *Colletotrichum*; *Diplocarpon*; *Elsinoe*; *Diaporthe*; *Sphaerotheca*; *Cinula*, *Cercospora*; *Erysiphe*; *Sphaerotheca*; *Leveillula*; *Mycosphaerella*; *Phyllactinia*; *Gloesporium*; *Gymnosporangium*, *Leptotthrydium*, *Podosphaera*; *Gloedes*; *Cladosporium*; *Phomopsis*; *Phytopora*; *Phytophthora*; *Erysiphe*; *Fusarium*; *Verticillium*; *Glomerella*; *Drechslera*; *Bipolaris*; *Personospora*; *Phaeoisariopsis*; *Spaceloma*; *Pseudocercospora*; *Pseudoperonospora*; *Puccinia*; *Typhula*; *Pyricularia*; *Rhizoctonia*; *Stachosporium*; *Ucinula*; *Ustilago*; *Gaeumannomyces* and *Fusarium*, more preferred from the group consisting of the genera and species *Alternaria*, *Podosphaera*, *Sclerotinia*, *Physalospora* such as *Physalospora* canker, *Botrytis* species such as *Botrytis* cinerea, *Corynespora* such as *Corynespora* melonis; *Colletotrichum*; *Diplocarpon* such as *Diplocarpon* rosae; *Elsinoe* such as *Elsinoe* fawcetti, *Diaporthe* such as *Diaporthe* citri; *Sphaerotheca*; *Cinula* such as *Cinula* neccata, *Cercospora*; *Erysiphe* such as *Erysiphe* cichoracearum and *Erysiphe* graminis; *Sphaerotheca* such as *Sphaerotheca* fuliginea; *Leveillula* such as *Leveillula* taurica; *Mycosphaerella*; *Phyllactinia* such as *Phyllactinia* kakicola; *Gloesporium* such as *Gloesporium* kaki; *Gymnosporangium* such as *Gymnosporangium* yamadae, *Leptotthrydium* such as *Leptotthrydium* pomi, *Podosphaera* such as *Podosphaera* leucotricha; *Gloedes* such as *Gloedes* pomigena; *Cladosporium* such as *Cladosporium* carpophilum; *Phomopsis*; *Phytopora*; *Phytophthora* such as *Phytophthora* infestans; *Verticillium*; *Glomerella* such as *Glomerella* cingulata; *Drechslera*; *Bipolaris*; *Personospora*; *Phaeoisariopsis* such as *Phaeoisariopsis* vitis; *Spaceloma* such as *Space-*

- loma ampelina; Pseudocercospora such as Pseudocercospora herpotrichoides; Pseudoperonospora; Puccinia; Typhula; Pyricularia such as Pyricularia oryzae; Rhizoctonia; Stachosporium such as Stachosporium nodorum; Uncinula such as Uncinula necator;
- 5 Ustilago; Gaeumannomyces species such as Gaeumannomyces graminis and Fusarium such as Fusarium dimerium, Fusarium merismoides, Fusarium lateritium, Fusarium decemcellulare, Fusarium poae, Fusarium tricinctum, Fusarium sporotrichioides, Fusarium chlamydosporum, Fusarium moniliforme, Fusarium proliferatum, Fusarium anthophilum, Fusarium subglutinans, Fusarium nygamai, Fusarium oxysporum,
- 10 Fusarium solani, Fusarium culmorum, Fusarium sambucinum, Fusarium crookwellense, Fusarium avenaceum ssp. avenaceum, Fusarium avenaceum ssp. aywerte, Fusarium avenaceum ssp. nurragi, Fusarium heterosporum, Fusarium acuminatum ssp. acuminatum, Fusarium acuminatum ssp. armeniacum, Fusarium longipes, Fusarium compactum, Fusarium equiseti, Fusarium scripi, Fusarium polyphialidicum, Fusarium semitectum and Fusarium beomiforme and especially preferred from the genera Fusarium such as Fusarium graminearum, most preferred from the group consisting of the genera and species Fusarium,
- 20 Fusarium dimerium, Fusarium merismoides, Fusarium lateritium, Fusarium decemcellulare, Fusarium poae, Fusarium tricinctum, Fusarium sporotrichioides, Fusarium chlamydosporum, Fusarium moniliforme, Fusarium proliferatum, Fusarium anthophilum, Fusarium subglutinans, Fusarium nygamai, Fusarium oxysporum, Fusarium solani, Fusarium culmorum, Fusarium sambucinum, Fusarium crookwellense, Fusarium avenaceum ssp. avenaceum, Fusarium avenaceum ssp. aywerte, Fusarium avenaceum ssp. nurragi, Fusarium heterosporum, Fusarium acuminatum ssp. acuminatum, Fusarium acuminatum ssp. armeniacum, Fusarium longipes, Fusarium compactum, Fusarium
- 25 equiseti, Fusarium scripi, Fusarium polyphialidicum, Fusarium semitectum and Fusarium beomiforme, wherein Fusarium graminearum is most preferred.

- Preferred non-phytopathogenic filamentous fungi are fungi of the
- 35 group consisting of the genera Neurospora such as Neurospora crassa, Aspergillus such as Aspergillus parasiticus, Aspergillus nidulans, Aspergillus niger and Wangiella such as Wangiella dermatidis.
- 40 The term "comprising" means that the nucleic acid sequence according to the invention can be flanked by additional nucleic acid sequences that have on the 5' end a sequence length of at least 1000 bp and preferably at least 500 bp, more preferably at least 100bp, most preferably at least 50bp and on the 3' a sequence length of at least 1000 bp and preferably at least 500 bp,
- 45

more preferably at least 100 bp and most preferably at least 50bp.

5 "Functional equivalents" in the present context describe nucleic acid sequences which hybridize under standard conditions with the nucleic acid sequence or portions of the nucleic acid sequence having the function of the a selection marker.

10 It is advantageous to use short oligonucleotides of a length of 10-50 bp, preferably 15-40 bp, for example of the conserved or other regions, which can be determined via comparisons with other related genes in a manner known to the skilled worker for the hybridization. Alternatively, it is also possible to use longer  
15 fragments of the nucleic acids according to the invention or the complete sequences for the hybridization. These standard conditions vary depending on the nucleic acid used, viz. oligonucleotide, longer fragment or complete sequence, or depending on which type of nucleic acid, viz. DNA or RNA, is being used for the hybridization. Thus, for example, the melting temperatures for  
20 DNA:DNA hybrids are approx. 10°C lower than those of DNA:RNA hybrids of equal length.

Standard conditions are understood to mean, depending on the nucleic acid, for example temperatures between 42 and 58°C in an  
25 aqueous buffer solution with a concentration of between 0.1 and 5 x SSC (1 x SSC = 0.15 M NaCl, 15 mM sodium citrate, pH 7.2) or additionally in the presence of 50% formamide such as, for example, 42°C in 5 x SSC, 50% formamide. The hybridization conditions  
30 for DNA:DNA hybrids are advantageously 0.1 x SSC and temperatures of between approximately 20°C and 45°C, preferably between approximately 30°C and 45°C. The hybridization conditions for DNA:RNA hybrids are advantageously 0.1 x SSC and temperatures of between approximately 30°C and 55°C, preferably between approximately 45°C  
35 and 55°C. These temperatures stated for the hybridization are melting temperature values which have been calculated by way of example for a nucleic acid with a length of approx. 100 nucleotides and a G + C content of 50% in the absence of formamide. The experimental conditions for DNA hybridization are described in  
40 specialist textbooks of genetics such as, for example, Sambrook et al., "Molecular Cloning", Cold Spring Harbor Laboratory, 1989 and can be calculated using formulae known to the skilled worker, for example as a function of the length of the nucleic acids, the type of the hybrids or the G + C content. The skilled worker can  
45 find more information on hybridization in the following textbooks: Ausubel et al. (eds), 1985, Current Protocols in Molecular Biology, John Wiley & Sons, New York; Hames and Higgins (eds),



## 15

1985, Nucleic Acids Hybridization: A Practical Approach, IRL Press at Oxford University Press, Oxford; Brown (ed), 1991, Essential Molecular Biology: A Practical Approach, IRL Press at Oxford University Press, Oxford.

5

A functional equivalent is furthermore also understood to mean, in particular, natural or artificial mutations of the relevant nucleic acid sequences of the polyketide synthetase (PKS) as set forth in SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4 or  
10 SEQ ID NO:5 and its homologs from other organisms, wherein mutations comprise substitutions, additions, deletions, inversions or insertions of one or more nucleotide residues. This may also lead to a modification of the corresponding amino acid sequence of the PKS by substitution, insertion or deletion of one or more amino  
15 acids.

Thus, the scope of the present invention also extends to, for example, those nucleotide sequences which are obtained by modification of the nucleic acid sequence of the selection marker described by SEQ ID NO:1 or by SEQ ID NO:2 or SEQ ID NO:3, SEQ ID  
20 NO:4, SEQ ID NO:5 respectively. The purpose of such a modification can be, for example, the insertion of further cleavage sites for restriction enzymes, the removal of excess DNA, or the addition of further sequences. Said nucleic acid sequences should  
25 still maintain the desired function as marker for targeted transformation, despite the deviating nucleic acid sequence.

The term "identity" or "homology" between two nucleic acid sequences or polypeptide sequences is defined by the identity of  
30 the nucleic acid sequence/polypeptide sequence by in each case the entire sequence length, which is calculated by alignment with the aid of the program algorithm GAP (Wisconsin Package Version 10.0, University of Wisconsin, Genetics Computer Group (GCG),  
35 Madison, USA), setting the following parameters:

Gap Weight: 8

Length Weight: 4

Average Match: 2,912

Average Mismatch:-2,003

40

The term homology when used herein is the same as the term identity.

45

Functional equivalents thus encompass naturally occurring variants of the sequences described herein, and also artificial, for example chemically synthesized, nucleic acid sequences adapted to

the codon usage, or the amino acid sequences derived therefrom.

Moreover, SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4 or  
SEQ ID NO:5, nucleic acid sequences derived from the amino acid  
5 sequence SEQ ID NO:6 by back translation or parts of the afore-  
mentioned nucleic acid sequences can be used for the detection  
and isolation of functional equivalents of other fungi on the ba-  
sis of sequence identities. In this context, part or all of the  
sequence of the SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID  
10 NO:4 or SEQ ID NO:5 or nucleic acid sequences derived from the  
amino acid sequence SEQ ID NO:6 by back translation can be used  
as probe (e.g. hybridization probe) for screening in a genomic  
library or a cDNA library of the fungal species in question or in  
a computer search for sequences of functional equivalents in  
15 electronic databases. Especially for computer search for se-  
quences of functional equivalents in electronic databases, the  
amino acid sequence SEQ ID NO:6 or parts of the amino acid se-  
quence SEQ ID NO:6 are useful.

20 For the preparation of hybridization probes, SEQ ID NO:1, SEQ ID  
NO:2, SEQ ID NO:3, SEQ ID NO:4 or SEQ ID NO:5 or parts of the  
aforementioned nucleic acid sequences can be used. The prepara-  
tion of these probes and the experimental procedure are known.  
For example, this can be effected via the tailor-made preparation  
25 of radioactive or nonradioactive probes by means of PCR and the  
use of suitably labeled oligonucleotides, followed by hybridiza-  
tion experiments. The technologies required for this purpose are  
given, for example, in T. Maniatis, E.F. Fritsch and J. Sambrook,  
"Molecular Cloning: A Laboratory Manual", Cold Spring Harbor Lab-  
30 oratory, Cold Spring Harbor, NY (1989). The probes in question  
can furthermore be modified by standard technology (lit. SDM or  
random mutagenesis) in such a way that they can be employed for  
other purposes, for example as probe which hybridizes specifi-  
cally with mRNA and the corresponding coding sequences in order  
35 to analyze the corresponding sequences in other organisms.

Furthermore, the cDNA could be used to engineer recombinant mi-  
croorganisms to produce polyketide agents of pharmaceutical or  
agricultural interest as described by Pfeifer et al. (Pfeifer  
40 BA, Admiraal SJ, Gramajo H, Cane DE, Khosla C., Science 2001 Mar  
2;291(5509):1790-2). Thus, the present invention also comprises  
polypeptides with the biological activity of a polyketide syn-  
thetase encoded by a nucleic acid sequence comprising

45 i. a nucleic acid sequence shown in SEQ ID NO:5 or

- ii. a nucleic acid sequence which, owing to the degeneracy of the genetic code, can be deduced from the amino acid sequence shown in SEQ ID NO:6 by back translation; or
- 5 iii. nucleic acid sequence which is encoded by a functional analogue of an amino acid sequence that has at least an identity of 50% with the SEQ ID NO:6.

The term "functional analogues" describes nucleic acid sequences which are capable of bringing about the expression, in a filamentous fungi, of a polypeptide with the biological activity of polyketide synthetase and which can be deduced from an amino acid sequence by back translation which has a defined degree of identity with SEQ ID NO:6. The functional analogues set forth in iii) have at least an identity of 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64% or 65% or preferably of 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78% or 79% more preferably of 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89% or 90% most preferably of 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98% or 99% with the SEQ ID NO:6.

Thus, the present invention also encompasses, for example, those nucleotide sequences which are obtained by modification of the abovementioned nucleic acid sequences. For example, such modifications can be generated by techniques with which the skilled worker is familiar, such as "site directed mutagenesis", "error prone PCR", "DNA shuffling" (Nature 370, 1994, pp.389-391) or "staggered extension process" (Nature Biotechnol. 16, 1998, pp.258-261). The purpose of such a modification can be, for example, the insertion of further cleavage sites for restriction enzymes, the removal of DNA in order to truncate the sequence, the substitution of nucleotides in order to optimize the codons, or the addition of further sequences. Proteins which are encoded via modified nucleic acid sequences must retain the desired functions despite a deviating nucleic acid sequence.

Functional analogues thus comprise naturally occurring variants of the herein-described sequences and artificial nucleic acid sequences, for example those which have been obtained by chemical synthesis and which are adapted to the codon usage, and also the amino acid sequences derived from them.

As explained above, also the expression cassette or the vector comprising a PKS encoding nucleic acid sequence may comprise at least an additional selection marker, preferably the hygromycin resistance gene so that in a particular preferred embodiment, the selection of the successfully transformed filamentous fungi can be

## 18

carried out by hygromycin resistance of successfully transformed clones and by the presence of pigment (color) of successfully transformed clones. Most preferably, the vector comprising the PKS encoding nucleic acid sequence is a vector according to the invention comprising a PKS encoding nucleic acid sequence. In addition to the aforementioned selection method homologous recombination can be confirmed by PCR based on oligonucleotides preferably derived from the vector sequence flanking the 5 and 3 region of the gene to be inserted. Specific examples of these primers are given in the examples.

The invention furthermore relates to the use of polyketide synthetase encoding nucleic acid sequences as marker for targeted transformation in filamentous fungi.

Preferably, the present invention comprises the use of a nucleic acid sequence comprising

- a) a nucleic acid sequence encoding a polyketide synthetase; or
- b) parts of the nucleic acid sequence as defined in i., ii. or iii. consisting of at least 300bp

for transformation of filamentous fungi.

Preferred is the use of a nucleic acid sequence as marker for targeted transformation in filamentous fungi said nucleic acid comprising

- i. a nucleic acid sequence shown in SEQ ID NO:1 or SEQ ID NO:2; or
- ii. parts of the nucleic acid sequence as defined in i. consisting of at least 300bp, preferably at least 400bp, more preferably at least 450bp and most preferably at least 500bp;

Equally preferred is the use of a nucleic acid sequence as marker for targeted transformation in filamentous fungi, said nucleic acid comprising

- iii. a nucleic acid sequence shown in SEQ ID NO:3, SEQ ID NO:4 or SEQ ID NO:5; or
- iv. a nucleic acid sequence which, owing to the degeneracy of the genetic code, can be deduced from the amino acid sequence shown in SEQ ID NO:6 by back translation; or

## 19

- v. a functional equivalent of the nucleic acid sequence set forth in i) or iii) which is encoded by an amino acid sequence that has at least an identity of 40% with the SEQ ID NO:6; or
- 5
- vi. a nucleic acid sequence shown in SEQ ID NO:9 or SEQ ID NO:11;
- vii. a nucleic acid sequence which, owing to the degeneracy of the genetic code, can be deduced from the amino acid sequence shown in SEQ ID NO:10, SEQ ID NO:12 or SEQ ID NO:13 by back translation; or
- 10
- viii. parts of the nucleic acid sequence as defined in iii., iv, v., vi or vii. consisting of at least 300bp, preferably at least 400bp, more preferably at least 450bp and most preferably at least 500bp; or
- 15
- ix. parts of the nucleic acid sequence as defined in iii., iv, v., vi or vii. consisting of at least 300bp, preferably at least 400bp, more preferably at least 450bp, and most preferably at least 500bp comprising
- 20
- a) a nucleic acid sequence shown in SEQ ID NO:7; or
- 25
- b) a nucleic acid sequence which, owing to the degeneracy of the genetic code, can be deduced from the amino acid sequence shown in SEQ ID NO:8 by back translation; or
- 30
- c) a functional equivalent of a nucleic acid sequence set forth in a) which is encoded by an amino acid sequence that has at least an identity of 68% with the SEQ ID NO:8.

35 The nucleic acid sequences according to i. to ix encode for a polypeptide with the biological function of a polyketide synthetase or for a fragment of the aforementioned polypeptide.

Under the aforementioned sequences, the nucleic acid sequences according to i., ii., iii., iv., v. as well as parts of the aforementioned nucleic acid sequence consisting of at least 300bp, preferably at least 400bp, more preferably at least 450bp, most preferably at least 500bp are preferred. Those parts are preferably those set forth in ix.

## 20

Preferred phytopathogenic and non-phytopathogenic filamentous fungi are those mentioned above. The aforementioned nucleic acid sequences are hereinbelow also termed "PKS marker". Preferably, the term "PKS marker" designates nucleic acid sequences according to i., ii., iii., iv., v. as well as parts of the aforementioned nucleic acid sequence consisting of at least 300bp, preferably at least 400bp, more preferably at least 450bp, and most preferably at least 500bp are preferred.

- 10 The functional equivalents of the nucleic acid sequence set forth in iv. can be deduced from a functional equivalent of the amino acid sequence shown in SEQ ID NO:6 by back translation having at least an identity of 40%, 41%, 42%, 43%, 44%, 45%, 46%, 47%, 48% or 49% preferably of 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78% or 79% more preferably of 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89% or 90% and most preferably of 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98% or 99% with the SEQ ID NO:6.

20

- The functional equivalents of the nucleic acid sequence set forth in ix.c) can be deduced from a functional equivalent of the amino acid sequence shown in SEQ ID NO:8 by back translation having at least an identity of 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77% or 78% preferably of 79%, 80%, 81%, 82%, 83%, 84% or 85% more preferably of 86%, 87%, 88%, 89% or 90% most preferably of 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98% or 99% with the SEQ ID NO:8.

- The use of a PKS marker for targeted transformation of filamentous fungi can be based on significant reduction in the amount of polyketide synthetase which is present in a filamentous fungi. A reduction in the amount the polyketide synthetase means that the amount of polypeptide is reduced via recombinant methods.

- Reduction via recombinant methods can involve "antisense techniques", which describes a technology for the suppression (reduction) of expression of polyketide synthetase, where a PKS marker is transformed into the respective filamentous fungi in "antisense" orientation under the control of a suitable promoter. This method is used preferably for *Aspergillus* species, and more preferably for *Aspergillus nidulans*. The technologies required herefore are well known by the skilled artisan (for example see Bautista et al., Appl. Environ. Microbiol. 2000; 66(10) 4579-81). Suitable vectors therefore comprise an expression cassette comprising

## 21

- a) a promotor sequence in functional linkage with a PKS marker in antisense orientation; and optionally
- b) further genetic control sequences functionally linked to a nucleic acid sequence according to a).

The afore-mentioned expression cassette is hereinbelow termed as "PKS Marker expression cassette".

- 10 The term "expression cassette" can be defined as follows: An expression cassette comprises a nucleic acid sequence which should be expressed, linked functionally to at least one genetic control element, such as a promoter, and, advantageously, a further control element, such as a terminator. Examples of suitable promoters and terminators are given above. The nucleic acid sequence of the expression cassette can be, for example, a genomic or complementary DNA sequence or an RNA sequence, and the semisynthetic or fully synthetic analogs thereof. These sequences can exist in linear or circular form, extrachromosomally or integrated into the genome. The nucleic acid sequences in question can be synthesized or obtained naturally or comprise a mixture of synthetic and natural DNA components, and consist of a variety of heterologous gene segments from various organisms.
- 25 Artificial nucleic acid sequences are also suitable in this context as long as they make possible the expression, in a cell or organism, of a polypeptide encoded by a nucleic acid sequence according to the invention and having the biological activity of a polyketide synthetase. For example, synthetic nucleotide sequences can be generated which have been optimized with regard to the codon usage of the organisms to be transformed.

- All of the abovementioned nucleotide sequences can be generated from the nucleotide units by chemical synthesis in the manner known per se, for example by fragment condensation of individual, overlapping complementary nucleotide units of the double helix. Oligonucleotides can be synthesized chemically for example in the manner known per se using the phosphoamidite method (Voet, Voet, 2nd Edition, Wiley Press New York, pp. 896-897). When preparing an expression cassette, various DNA fragments can be manipulated in such a way that a nucleotide sequence with the correct direction of reading and the correct reading frame is obtained. The nucleic acid fragments are linked to each other via general cloning techniques as are described, for example, in T. Maniatis, E.F. Fritsch and J. Sambrook, "Molecular Cloning: A Laboratory Manual", Cold Spring Harbor Laboratory, Cold Spring Harbor, NY (1989) and in T.J. Silhavy, M.L. Berman and L.W. Enquist, Experi-

ments with Gene Fusions, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY (1984) and in Ausubel, F.M. et al., "Current Protocols in Molecular Biology", Greene Publishing Assoc. and Wiley-Interscience (1994).

5

The term "genetic control element" describes sequences which have an effect on the transcription and, if appropriate, translation of the nucleic acids according to the invention in prokaryotic or eukaryotic organisms. Examples are terminators. Examples of suitable terminators are given above. In addition to the aforementioned control sequences, or instead of these sequences, the natural regulation of these sequences may still be present before the actual structural genes and may, if appropriate, have been modified genetically in such a way that the natural regulation has been switched off and the expression of the target gene has been modified, that is to say increased or reduced. The choice of the control sequence depends on the host organism or starting organism. Genetic control sequences furthermore also comprise the 5'-untranslated region, introns or the noncoding 3' region of genes. Control sequences are furthermore understood as meaning those which make possible a homologous recombination or insertion into the genome of a host organism or which permit the removal from the genome. Genetic control sequences also comprise further promoters, promoter elements or minimal promoters.

25

The transcription of the PKS marker leads to suppression of the transcription of the natural polyketide synthetase gene, which can be detected by loss of color of the transformed fungi relative to the respective wild-type strain.

30

In a preferred embodiment, the reduction via recombinant methods is based on a gene knock out of the polyketide synthetase gene using either an expression cassette additionally comprising the PKS marker or a vector comprising the PKS marker in the respective filamentous fungi. Disruption of the PKS marker will lead to a loss of color.

35

Preferred phytopathogenic filamentous fungi are those mentioned above. Preferred non-phytopathogenic filamentous fungi are fungi of the group consisting of the genera *Aspergillus* such as *Aspergillus parasiticus*, *Aspergillus nidulans* and *Wangiella* such as *Wangiella dermatidis*.

40

In this connection, the selection of the functional equivalent for the use as marker gene depends on the fungi to be transformed. By preference, the polyketide synthetase fragment has an identity of at least 80%, preferably at least 81%, 82%, 83%, 84%,

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## 23

85%, 86%, 87%, 88%, 89%, 90%, and especially preferably at least 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% identity with the polyketide synthetase of the fungi to be transformed.

5 For example, for transformation of *Fusarium graminearum*, a nucleic acid sequence can be selected comprising

i. a nucleic acid sequence shown in SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4 or SEQ ID NO:5; or

10

ii. a nucleic acid sequence that has at least an identity of 80% SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4 or SEQ ID NO:5; or

15 iii. parts of the nucleic acid sequence as defined in i. or ii. consisting of at least 300bp, preferably at least 400bp, more preferably at least 450bp, and most preferably at least 500bp.

20 iv. parts of the nucleic acid sequence as defined in i. or ii. consisting of at least 300bp, preferably at least 400bp, more preferably at least 450bp, and most preferably at least 500bp comprising

25 a) a nucleic acid sequence shown in SEQ ID NO:7; or

c) a nucleic acid sequence that has at least an identity of 80% with the SEQ ID NO:8.

30 As mentioned above, another embodiment of the present invention is plasmid vectors for targeted transformation of filamentous fungi comprising a PKS marker. These plasmid vectors are either vectors currently used for targeted transformation of filamentous fungi e.g. such as pAN7 (Punt et al, 1987 Gene 36:117-124) and  
35 other vectors that are well known by the skilled artisan or plasmid vectors according to the invention, preferably plasmid vectors according to the invention.

All of the above-mentioned vectors comprising the PKS marker are  
40 hereinbelow termed as "PKS vectors".

A PKS vector is also a vector, which comprises a PKS Marker-expression cassette.

45 All vectors according to the invention not comprising the PKS marker are hereinbelow termed as "non-PKS vectors".

## 24

The present invention furthermore encompasses a method for preparing mutated filamentous fungi, comprising the steps of transferring a non-PKS vector or a PKS vector into a filamentous fungi; and selecting clones of said filamentous fungi, which contain  
5 at least one genetic marker introduced by said plasmid vector.

The term filamentous fungi as well as preferred fungi for this purpose are those mentioned and defined above.

10 In a preferred embodiment, the method for preparing mutated filamentous fungi, comprising the following steps

a) transferring a PKS vector into a filamentous fungi; and

15 b) selecting successfully transformed filamentous fungi by the absence of color (pigment).

As explained above, the absence of color is based on significant reduction in the amount of polyketide synthetase (or in the polyketide syntethase activity or instability of polyketide syntethase mRNA, which is present in a filamentous fungi). The absence  
20 of color can be monitored for example by comparing the transformed fungi with the respective wild-type fungi of the same species.

25

If a PKS vector is transferred into a filamentous fungi, the disruption of the PKS gene leads to a loss of color (pigment) whereby the degree of transformation can be determined easily. Resulting transformants are white in contrast to the colored  
30 wild-type. Thus, the selection according to step b) is done by monitoring the absence of color (pigment) in the filamentous fungi. In a preferred embodiment, the absence of pigment is monitored by optical means.

35 Alternatively, the absence of color results from the reduction of the polyketide synthetase via antisense techniques. The absence of color hereby means a "reduction of color" or, preferably, loss of color. Absence of color means a reduction in color of at least 20%, preferably between 20 and 40%, more preferably between 40  
40 and 60%, especially preferably between 60 and 80% and most preferably between 80% and 100%.

In a more preferred embodiment, the PKS vector comprises at least an additional selection marker, preferably the hygromycin resistance gene. In a particular preferred embodiment, the selection of  
45 the successfully transformed filamentous fungi comprising a PKS vector can be carried out by hygromycin resistance of success-

## 25

fully transformed clones and by the absence of pigment of successfully transformed clones. Most preferably, the PKS vector is a vector according to the invention additionally comprising a PKS marker.

5

In a further embodiment of the invention, the selection of the successfully transformed filamentous fungi comprising a non-PKS vector can be carried out by hygromycin resistance of successfully transformed clones.

10

If a non-PKS vector is used, the vector is linearized by a restriction enzyme cutting in the nucleic acid sequence region of element d). Also nucleic acid sequences exceeding 2000 bp can be used what can be disadvantageous as mentioned above. If a PKS

15 vector is used, the plasmid vector is transferred into a filamentous fungi with the proviso that said vector is linearized by a restriction enzyme in PKS marker nucleic acid sequence. Unlike the non-PKS vectors, the nucleic acid sequence to be expressed recombinantly can also be smaller than 400bp.

20

In addition to the aforementioned selection methods set forth in step a) to c), homologous recombination can be confirmed by PCR based on oligonucleotides preferably derived from the vector sequence flanking the 5' and 3' regions of the gene to be inserted.

25 Specific examples of these primers are given in the examples..

The plasmid vector may be transferred into the filamentous fungi to be transformed by methods familiar to the skilled worker, preferably via protoplast preparation with driselase or driselase  
30 and glucanase as lytic enzyme.

The above-mentioned transformation methods can be also realized in a high throughput screening. Using high throughput screening, many different clones are obtained in parallel so that large numbers of successfully transformed clones of filamentous fungi can  
35 be quickly screened.

The term filamentous fungi as well as preferred fungi for this purpose are those mentioned and defined above.

40

Due to the convenience of the vector, the above-mentioned KO-plasmid preparation, fungi transformation and screening of the mutants can be at least partially automated so that the whole procedure can also be realized in a high throughput screening.

45 Using high throughput system for example for KO-plasmid preparation and DNA amplification by PCR to screen the recombinant mu-

tants, many different clones are obtained in parallel so that large numbers of transformants can be quickly screened.

Mutagenized filamentous fungi, obtainable according to a method 5 mentioned above, are further encompassed by the present invention.

In an alternative embodiment, the method of transforming filamentous fungi based on the use of polyketide synthetase as marker 10 for transformation comprises the following steps:

- a) providing a filamentous fungi characterized by the absence of color (pigment), in which the polyketide synthetase gene is modified in such a way that the polyketide synthetase cannot 15 be functionally expressed;
- b) transforming the filamentous fungi of step a) with a "sense expression cassette" or a vector comprising the aforementioned expression cassette; 20
- c) selecting successfully transformed filamentous fungi by the presence of pigment (color).

The nucleic acid sequence as defined in b) i to v. is herein below 25 termed as PKS encoding sequence.

The terms "expression cassette" and "genetic control elements" are explained above.

30 The "sense-expression cassette" set forth in step b) of the above-mentioned method comprises

- a) a promotor sequence in functional linkage with a nucleic acid sequence comprising 35
  - i. a nucleic acid sequence shown in SEQ ID NO:3, 4 or 5; or
  - ii. a nucleic acid sequence which, owing to the degeneracy of the genetic code, can be deduced from the amino acid sequence shown in SEQ ID NO:6 by back translation; or 40
  - iii. a functional equivalent of the nucleic acid sequence set forth in i) which is encoded by an amino acid sequence shown in SEQ ID NO:6 that has at least an identity of 40% 45 with the SEQ ID NO:6; or

iv. a nucleic acid sequence shown in SEQ ID NO:9 or SEQ ID NO: 11;

5 v. a nucleic acid sequence which, owing to the degeneracy of the genetic code, can be deduced from the amino acid sequence shown in SEQ ID NO:10, SEQ ID NO:12 or SEQ ID NO:13 by back translation;

and optionally

10

b) further genetic control sequences in functionally linked with a nucleic acid sequence according to a).

The expression cassette or vector comprises preferably a polyketide synthetase encoding nucleic acid sequence as set forth in b) i., ii. or iii..

Preferred phytopathogenic filamentous fungi are those mentioned above. Preferred non-phytopathogenic filamentous fungi are fungi of the group consisting of the genera *Aspergillus* such as *Aspergillus parasiticus*, *Aspergillus nidulans* and *Wangiella* such as *Wangiella dermatidis*.

The modification of the polyketide synthetase encoding sequence of the respective fungi can be done either by introduction of at least one mutation in the gene encoding a polyketide synthetase or disruption of the gene encoding a polyketide synthetase.

The term "disruption of the PKS marker" means that the PKS marker sequence is disrupted by introducing DNA comprising stop-codons in the PKS marker sequence e.g. by homologous recombination. The respective methods are well known by the skilled artisan.

The term "mutations" of nucleic acid sequences comprises substitutions, additions, deletions, inversions or insertions of one or more nucleotide residues, which have to bring about termination of translation of the corresponding amino acid sequence of the target protein by the substitution, insertion or deletion of one or more amino acids (e.g. a by frame-shift or introduction of stop codon or amendment of nucleic acid sequence). The respective methods are well known by the skilled artisan.

For example, the mutations are carried out in the flanking regions of exon and intron of a PKS gene. These regions can be determined easily by the skilled artisan. For example, in SEQ ID NO:3 the flanking regions between exon are at bp 1022/1023; bp 1067/1068, bp 1361/1362; bp 1067/1068; bp 1361/1362; bp

1067/1068; bp 1361/1362; bp 1416/1417; bp 2399/2400; bp  
2447/2448; bp 2675/2676; bp 2738/2739; bp 5744/5745; bp  
5792/5793; and/or bp 7205/7206 (Ende 6. exon bp 7205).

5 The term "functional analogues" is defined above describe, in the  
present context nucleic acid sequences which are capable of  
bringing about the expression, in a filamentous fungi, of a poly-  
peptide with the biological activity of polyketide synthetase and  
which can be deduced from an amino acid sequence by back transla-  
10 tion which has at least an identity of 40%, 41%, 42%, 43%, 44%,  
45%, 46%, 47%, 48% or 49%, preferably of 50%, 51%, 52%, 53%, 54%,  
55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%,  
68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78% or 79%,  
more preferably of 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%,  
15 89%, or 90% most preferred of 91%, 92%, 93%, 94%, 95%, 96%, 97%,  
98% or 99% with the SEQ ID NO:6.

As explained above, the plasmid vector may be transferred into  
the filamentous fungi to be transformed by methods familiar to  
20 the skilled worker, preferably via protoplast preparation with  
driselase or driselase and glucanase as lytic enzyme.

The above-mentioned transformation methods can also be realized  
in a high throughput screening. Using high throughput screening,  
25 many different clones are obtained in parallel so that large num-  
bers of successfully tranformed clones of filamentous fungi can  
be quickly screened.

30 The invention is now illustrated by the examples which follow,  
but is not limited thereto.

#### Examples

35 The recombinant methods on which the exemplary embodiments which  
follow are based are now described briefly:

#### A: General methods

40 Cloning methods such as, for example, restriction cleavages, DNA  
isolation, agarose gel electrophoresis, purification of DNA frag-  
ments, transfer of nucleic acids to nitrocellulose and nylon mem-  
branes, linking of DNA fragments, transformation of E. coli  
cells, bacterial cultures, sequence analysis of recombinant DNA  
45 and Southern and Western Blots were carried out as described by  
Sambrook et al., Cold Spring Harbor Laboratory Press (1989) and  
Ausubel, F.M. et al., Current Protocols in Molecular Biology,

Greene Publishing Assoc. and Wiley-Interscience (1994); ISBN 0-87969-309-6.

5 The bacterial strains used hereinbelow (E. coli DH5 or XL1 blue) were obtained from Life Technologies or Stratagene. The vectors were used for cloning. DSM:4527 can be used as *F. Graminearum* wild-type strain 8/1. Restriction maps of the vectors pUCmini-Hyg and PUCmini-Hyg TA are given in Fig 1 and 2.

10 B: Sequence analysis of recombinant DNA

Recombinant DNA molecules were sequenced using an ABI laser fluorescence DNA sequencer following the method of Sanger (Sanger et al., Proc. Natl. Acad. Sci. USA, 74, 5463-5467 (1977)). Fragments  
15 resulting from a polymerase chain reaction were sequenced and verified in order to avoid polymerase errors in constructs to be expressed.

20 C: Materials used

Unless otherwise specified in the text, all of the chemicals used were obtained in analytical grade quality from Fluka (Neu-Ulm), Merck (Darmstadt), Roth (Karlsruhe), Serva (Heidelberg) and Sigma  
25 (Deisenhofen). Solutions were prepared using pure pyrogen-free water, referred to in the following text as H<sub>2</sub>O, from a Milli-Q water system purification unit (Millipore, Eschborn). Restriction enzymes, DNA-modifying enzymes and molecular-biological kits were obtained from AGS (Heidelberg), Amersham (Brunswick), Biometra  
30 (Göttingen), Roche (Mannheim), Genomed (Bad Oeynhausen), New England Biolabs (Schwalbach/Taunus), Novagen (Madison, Wisconsin, USA), Perkin-Elmer (Weiterstadt), Pharmacia (Freiburg), Qiagen (Hilden) and Stratagene (Heidelberg). Unless otherwise specified, they were used following the manufacturer's instructions.

35

All of the media and buffers used for the genetic engineering experiments were sterilized either by filter sterilization or by heating in an autoclave.

40 In degenerated primer sequences, the following abbreviations are used:

A or T = "W";

G or C = "S";

T or C = "Y";

45

A or C = "M";

A or G = "R";

## Examples

Example 1 - Construction of pUCmini-Hyg and PUCmini-Hyg TA vector

5 A 2536 bp DNA fragment corresponding to the promoter of glycerol-3-phosphate dehydrogenase (GPD1) from *Cochliobolus heterotrophus* associated to the hygromycin B resistance gene from *Escherichia coli* was amplified by PCR with the oligonucleotides

10 P1 5' atgaagcttgggggtttgagggccaatggaacgaaactagtgtaccacttgacc 3' (SEQ ID NO 14); and

P2 5'gacagatctggcgccattcgccattcag 3' (SEQ ID NO 15)

15 using pGUS5 as template (Mönke, E. and Schäfer, W., 1993, Mol. Gen. Genet. 241: 73-80). The PCR is done using standard protocols; e.g. as described in Maniatis et al., Mol. Cloning.

The resulting DNA fragment was inserted in the plasmid pFDX3809  
20 (WO 01/38504) by the restriction site Hind III and Bgl II introduced by the oligonucleotides P1 and P2. The resulting plasmid pHygB serves as template for a further PCR, wherein the Oligonucleotides

25 ANK 518 5' ggaatcgggtcaatacactac 3' (SEQ ID NO 16)

ANK 519 5' tgtagatctctattcctttgccctcggacgagt 3' (SEQ ID NO 17)

are used to shorten the hygromycin B resistance gene specifically.  
30 The resulting PCR fragment comprising 575 bp of the 3' end of the hygromycin gene was inserted in the plasmid pHygB via the restriction sites Nde I/ Bgl II generating the plasmid pHygB-NOS.

A Hind III / Ssp I DNA fragment of 2019 bp containing the expression cassette GPD1 promoter, the hygromycin B resistance gene  
35 and the nopaline synthase terminator was isolated from pHygB-NOS and inserted in the pUCmini plasmid (= plasmid pFDX3809, see WO 01/38509) previously treated with EcoRI and HindIII restriction enzymes to give the plasmid pUCmini-Hyg; to do so , the EcoRI  
40 ends were made compatible with Ssp I by a fill-in treatment using the Klenow fragment of DNA polymerase I. A second version of pUCmini-Hyg, called pUCmini-Hyg-TA, was obtained by the insertion of the following adaptor in the NotI/AscI restriction sites of pUCmini-Hyg:

45

5' GGCCGCCACGGATATCTTGGCCAAAGAATTCCTGG 3' (SEQ ID NO 18)



## 31

3' CGGTGCCTATAGAACCGGTTTCTTAAGGACCGCGC 5' (SEQ ID NO 19)

The adaptor contains 2 XcmI restriction sites so that XcmI digest of pUCmini-Hyg-TA creates T-overhangs that permits direct cloning of PCR products made with the classical Taq-polymerases.

Example 2 - Construction of the PKS comprising vector "pUCmini-Hyg-PKS"

- 10 The nucleic acid sequence encoding PKS was amplified by PCR with degenerated primers

LC1 5'-GAY CCI MGI TTY TTY AAY ATG-3' (SEQ ID NO 20)

- 15 LC2c 5'-GTI CCI GTI CCR TGC ATY TC-3' (SEQ ID NO 21)

- based on the conserved amino acid sequence of the PKS gene sequences from *Aspergillus nidulans*, *Colletotrichum lagenarium*, *Penicillium patulum*, and *Aspergillus parasiticus* (Bingle et al., 1999) using genomic DNA of *Fusarium graminearum* as template. Thermal cycling parameters consisted of an initial denaturation at 94°C for 3 min followed by 34 cycles of 94°C for 1 min (denaturation), 55°C for 1 min (annealing), 72°C for 3 min (extension) and a final extension at 72°C for 10 min according to standard procedures. The resulting PCR product was cloned into the pGEM-T vector (Promega, Mannheim, Germany) to give the plasmid pGEM-T/PKS833 and sequenced. A 633 bp DNA fragment (2236bp to 2870bp of SEQ ID NO:1; corresponding to 2234bp to 2865bp of SEQ ID NO:3; set forth in SEQ ID NO:18) was amplified by PCR using the oligo- nucleotides

ANK593 5' ATAAGAATGCGGCCGCAATGGCCCTCGAAACAGC 3' (SEQ ID NO 22)

ANK594 5' AAATGGCGCGCCGCGCCCAGAATGACACC 3' (SEQ ID NO 23)

- 35 and cloned into the plasmid pUCmini-Hyg using the restriction site NotI and AscI present in the oligonucleotide sequences. The resulting plasmid pUCmini-Hyg-PKS is used for homologous recombination.

- 40 The flanking regions of the PKS DNA fragment were obtained by inverse PCR (Triglia T, Peterson MG, Kemp DJ, Nucleic Acids Res 1988 Aug 25;16(16):8186). Genomic DNA was treated with the restriction enzymes PstI, NcoI, or XhoI respectively. DNA was then self-ligated to get circular DNA molecule. The latter was used as template for the inverse PCR reaction using the primers

## 32

P1A: 5' TGCCACCTGTAGTCTGCAATCAG 3' (SEQ ID NO 24) and

P2A: 5' TGAATAACCCTGACAACTTCGCTG 3' (SEQ ID NO 25)

5 deduced from the polyketide synthetase (PKS) DNA fragment of the plasmid pGEM-T/PKS833 described above.

In a second step, the PCR product was reamplified with the nested primers

10

P1B: 5' CCAGGATCCGACTGCTCAG 3' (SEQ ID NO 26) and

P2B: 5' CTACATCGAGATGCACGGCAC 3' (SEQ ID NO 27)

15 (deduced from the PKS DNA fragment of the plasmid pGEM-T/PKS833), cloned into the pPCR-XL-TOPO vector (Invitrogen) and sequenced to get SEQ ID NO:1.

Identification of the genomic DNA Sequence

20

The remaining parts of the flanking regions were obtained by Tail-PCR (Liu YG, Whittier RF; Genomics 1995 Feb 10;25(3):674-81) using 9 arbitrary degenerated primers

25 FJM-tail-AD1 5'-NGT CGA SWG ANA WGA A-3' (SEQ ID NO 28),

FJM-tail-AD2 5'-GTN CGA SWC ANA WGT T-3' (SEQ ID NO 29),

FJM-tail-AD3 5'-WGT GNA GWA NCA NAG A-3' (SEQ ID NO 30),

30

FJM-tail-AD4 5'-NTC GAS TWT SGW GTT-3' (SEQ ID NO 31),

FJM-tail-AD6 5'-TGW GNA GWA NCA SAG A-3' (SEQ ID NO 32),

35 FJM-tail-AD7 5'-AGW GNA GWA NCA WAG G-3' (SEQ ID NO 33),

FJM-tail-AD8 5'-CAW CGI CNG AIA SGA A-3' (SEQ ID NO 34)  
and

40 FJM-tail-AD9 5'-TCS TIC GNA CIT WGG A-3' (SEQ ID NO 35),

coupled to the primer

45 TailPKS1c 5'-TTG TTA CTG GAG AGG TAA TGA AG-3' (SEQ ID NO 36)

specific for the 5' PKS flanking region deduced from SEQ ID NO:1,

or coupled to the primer

TailPKS2c 5'-TGA GAC AGA TCT CGC GAG CCC TC-3'. (SEQ ID NO 37)

- 5 specific for the 3' PKS flanking region deduced from SEQ ID NO:1. After subcloning and subsequent sequencing of the PCR products SEQ ID NO:3 was obtained.

#### Identification of the cDNA Sequence of Polyketide Synthetase

10

The PKS cDNA sequence was obtained by RT-PCR with a crude RNA preparation from *Fusarium graminearum* and various primers deduced from the genomic sequence. This was done according the classical methods (Ausubel, F.M. et al., Current Protocols in

- 15 Molecular Biology, Greene Publishing Assoc. and Wiley-Interscience (1994); ISBN 0-87969-309-6). Alignment of cDNA and genomic PKS sequences permits to be identified precisely the location of introns in the genomic sequence.

#### 20 Example 3 Transformation of *F. graminearum*

50 ml of CM-medium (Leach et al., 1982, J. Gen. Microbiol. 128: 1719-1729) were inoculated with approximately  $10^5$  conidia, and incubated for 2 days at 28°C, 140 rpm. Resulting hyphae were homoge-

- 25 nized in a Warring-Blender; 200 ml CM were inoculated with 10 ml hyphal suspension, and incubated overnight at 24°C. Mycel were trapped on a sterile filter, and washed two times with sterile water. 2 g of the hyphae were resuspended in 20 ml Driselase/Glucanase (InterspeX Products, San Mateo, USA; 5% / 3% in 700 mM  
30 NaCl, pH 5.6), and digested 2½ to 3 h at 28°C, 75 rpm. Undigested hyphal were removed from the protoplast suspension by filtration through gauze and Nybold membrane (50 µm pore size). The protoplast suspensions were combined with 700 mM NaCl and again passed through the gauze and the Nybold membrane. The protoplasts were  
35 pelleted by centrifugation (1300 x g) in a swing-out rotor and washed two times with ice-cold NaCl 700 mM and centrifuged (830 x g). Then the protoplasts were resuspended in STC (0.8 M sorbitol, 50 mM Tris-HCl pH 8.0, 50 mM CaCl<sub>2</sub>) and stored on ice until transformation (maximum 1 week).

40

For transformation, protoplasts were resuspended in 4 parts STC and 1 part SPTC (0.8 M sorbitol, 40% polyethylene glycol 4000, 50 mM Tris-HCl pH 8.0, 50 mM CaCl<sub>2</sub>) at a concentration of  $0.5-2 \times 10^8$ /ml; 30 µg of the pUCmini-Hyg-PKS plasmid DNA linearized with

- 45 the Eco47III restriction site inside the PKS fragment and 5 µl heparin (5 mg/ml in STC) were added to 100 µl of the protoplast suspension in 10 ml tubes. After mixing, samples were incubated

## 34

- on ice for 30 min. 1 ml SPTC was mixed with the suspension and incubated at room temperature for 20 min. Protoplasts were mixed gently into 200 ml regeneration medium ( 0.1% (w/v) yeast extract, 0.1% (w/v) caseinhydrolysate, 34.2% (w/v) sucrose, 1.6% (w/v) granulated agar) at 43°C and spread on 94 mm plates (20 ml per plate). The plates were incubated at 28°C. After 12-24 h, the plates were overlaid with 10 ml per plate water based selective medium (16g/l granulated agar, 100mg/l hygromycin and further incubated at 28°C until transformants were obtained, which were transferred to fresh CM-Hyg-plates (consisting of CM-media, 100 µg/ml hygromycin and 2% (w/v) agar. The transformants were isolated by single spore isolation. For generation of conidia, the transformants were cultivated on SNA plates (Nirenberg, 1981, Canadian J. Botany 59: 1599-1609) under UV-light 7-14 days at 18°C. Dilutions of conidia were plated on CM-Hyg plates, and single colonies were transferred from these plates to fresh CM-Hyg plates.

## Example 4 Southernblot analysis

- Genomic DNA was isolated from frozen hyphal material using the Puregene Genomic DNA Isolation Kit (Gentra Systems, Minneapolis USA) and digested for 6 h with *Nru*I restriction enzyme. The genomic DNA was separated by electrophoresis on a 1% (w/v) agarose gel and blotted onto a nylon membrane (Hybond NX; Amersham Pharmacia Biotech, Buckinghamshire, England). A digoxigenin labeled probe was generated by PCR based on specific primers PKS forward 5'-GCG CTT GAG ATG GCT AGT ATC G-3' and and PKS reverse 5'-GTG CCG TGC ATC TCG ATG TAG-3' using pGEM-T/PKS833 as template and digoxigenin labeled dUTPs by PCR reaction according to the recommendation of the manufacturer (Roche Diagnostics GmbH, Mannheim). PCR conditions were 94°C for 3 min (initial denaturation) followed by 30 cycles of 94°C for 30 sec (denaturation), 55°C for 45 sec (annealing), 72°C for 1 min (extension) and a final extension at 72°C for 10 min. The non-radioactive hybridization and the detection were done under highly stringent conditions as described in Roche Molecular Biochemicals DIG Application Manual for Filter Hybridization (Roche Diagnostics GmbH, Mannheim).
- To confirm the insertion of the vector construct into the PKS locus in comparison with the wild type gene, primers
- EF-PKS 5' atgtctccaaaggaagctgagc 3' (SEQ ID NO 38); and
- ER-PKS 5'tcgagtgatggatactgcttcg 3' (SEQ ID NO 39)

## 35

are constructed based on the PKS DNA sequence from the plasmid pGEM-T/PKS833; four universal primers are constructed, wherein

Lac 92 5' cggctacactagaaggacagtatttggtta 3' (SEQ ID NO 40)

5

Lac 93 5' gtcaggcaactatggatgaacgaaatagac 3' (SEQ ID NO 41)

Lac 94 5' acccatctcataaataacgctcatgc 3' (SEQ ID NO 42); and

10 Lac 95 5' caactctatcagagccttggttga 3' (SEQ ID NO 43)

permit amplification of a 412 bp DNA fragment of the hygromycin cassette.

- 15 PCR reactions were conducted in classical conditions: 94°C for 3 min (initial denaturation) followed by 30 cycles of 94°C 60 sec (denaturation), 55°C for 90 sec (annealing), 72°C for 90 sec (extension) and a final extension at 72°C for 10 min. Six (6) recombinant clones resistant to hygromycine were analyzed
- 20 by PCR using the primer set Lac 94 / Lac 95 specific for the hygromycin resistance gene. All the mutants were found to present the expected DNA fragment of 412 bp, indicating the integration of the plasmid pUCmini-Hyg-PKS in the genome.
- 25 A 712 bp corresponding to the PKS gene could be amplified with the primer set EF-PKS/ER-PKS mentioned above using genomic DNA from a wild type strain; however no PCR fragments were amplified with genomic DNA from the recombinant clones indicating that the PKS gene is disrupted by the insertion of pUCmini-Hyg-PKS. This
- 30 was confirmed by PCR amplification EF-PKS combined with Lac 93 (hybridizing to the plasmid backbone near Not I restriction site) and ER-PKS combined with Lac 92 (hybridizing to the plasmid backbone near Asc I restriction site). In both cases, DNA fragments of about 600 bp were amplified for the recombinant clones but not
- 35 for the wild type strain (WT). Altogether, the PCR analysis using the different primer sets proves that the plasmid pUCmini-Hyg-PKS was targeted specifically in the PKS locus by homologous recombination. This process permits the PKS gene to be disrupted since the recombinant mutants were found to lack the typical pig-
- 40 mentation (purple) of the wildtype strain.

Example 5 functional expression of Green Fluorescent Protein (GFP) in *Fusarium graminearum*

45 A) Plasmid construction

## 36

In a first step, a 67bp DNA fragment encoding the peptide leader of the first 23 amino acids from N-terminus of the yeast ARH1 (SwissProt;P48360) was amplified by PCR using the primers

5 Lac 80 5' cccgaattcatgagctttgttcaaataagg 3' (SEQ ID NO 44) and

Lac 81 5' ttattctagattttccatgggaatggatacagtcttacg 3' (SEQ ID NO 45)

- 10 In a second step, a 734 bp DNA fragment encoding the Green Fluorescent Protein (GFP) was amplified by PCR using the plasmid pEGFP-N2 (Genbank; U57608) and the primers

15 Lac 84 5' cgccaccatggtgagcaagggcgaggagctgtt 3' (SEQ ID NO 46) and

Lac 85 5' tatgatctagagtcgcggccgctttacttgtacagctcg 3' (SEQ ID NO 47).

- 20 The PCR products were assembled in frame with the Nco I restriction sites present in the oligonucleotides Lac 81 and Lac 84 and cloned in the expression plasmid pYes2 (Invitrogen) using the restriction sites EcoRI and Xba I present in the oligonucleotides Lac 80 and Lac 85, respectively. In the resulting plasmid pLAC7, 25 the recombinant gene encoding GFP is under the control of the galactose (Gal 1) promoter and cytochrome C1 terminator.

- A 2892 bp DNA fragment containing the GFP expression cassette was isolated from pLac7 using the restriction sites Nae I and Bsa I 30 and cloned in the plasmid pUCmini-Hyg-PKS (see example 2). To do so, pUCmini-Hyg-PKS was firstly cut by Asc I and filled in according to classical method then treated with Bsa I. The resulting plasmid pUCmini-Hyg-PKS-GFP contains all genetic elements permitting the production of recombinant GFP in *Fusarium graminearum*. 35

B) Transformation of *Fusarium graminearum* with pUCmini-Hyg-PKS-GFP and analysis of transformants

- 40 The transformation was done as described in example 3, wherein pUCmini-Hyg-PKS-GFP was linearized with EcoR47III. The correct integration of the plasmid in the PKS locus was observed after single conidiation by the absence of pigmentation of the recombinant mutants.

45

37

In addition, the integration was confirmed by PCR as described in example 4 using the following primer combinations EF-PKS (see example 4; SEQ ID NO:38) and ER-PKS (see example 4; SEQ ID NO:39), whereby no amplification were observed since the gene PKS is disrupted, whereas wild type strain or unspecific mutants presented a 714 bp DNA fragment corresponding to the expected PKS DNA fragment.

Using the primer combination EF-PKS (see example 4; SEQ ID NO:38) and

Lac 211 5' gcttctaataccgtactagtgatca 3' (SEQ ID NO 48)

the amplification of a 835 bp DNA corresponding to the 5' end plasmid integration in the PKS locus of the mutants was observed. No DNA fragment was amplified for the wildtype strain due the absence of the DNA sequence complementary to Lac211.

The primer combination

ANK 458 5' ctttgatcttttctacgggggtctga 3' (SEQ ID NO 49) and

ER-PKS (see example 4; SEQ ID NO:39) led to the amplification of a 718 bp DNA corresponding to the 3' end plasmid integration in the PKS locus of the mutants. No DNA fragment was amplified for the wildtype strain due the absence of the DNA sequence complementary to ANK 458.

C) Detection of the production of GFP

The recombinant mutants were grown for a few days in CM-Hyg medium as described in example 3 except for glucose which was replaced by galactose as a carbon source. The fluorescence of GFP was detected using the polarstar spectrophotometer (Firma BMG; Ex: 385nm and Em: 520nm). In these conditions fluorescence was observed for the strains which showed integration of the plasmid whereas no fluorescence was observed for the wildtype strains.

Brief description of the figures

Figure 1: Map of pUCmini-Hyg

Figure 1: Map of PUCmini-Hyg TA

## Claims

1. A plasmid vector for targeted transformation of filamentous  
5 fungi comprising
  - a) an origin of replication for a host organism not originating from the filamentous fungi to be transformed;
  - 10 b) a selection marker for a host organism not originating from the filamentous fungi;
  - c) a promotor facilitating recombinant expression in fungi that is functionally linked to the coding region of the  
15 hygromycin resistance gene which is functionally linked to a terminator which facilitates transcription termination in filamentous fungi;
- wherein the overall size of the elements a), b) and c) does  
20 not exceed 4500 bp; and
  - d) a nucleic acid sequence which is homologous to nucleic acid sequences of the filamentous fungi to be transformed and makes homologous recombination in the filamentous  
25 fungi to be transformed possible.
2. A plasmid vector as claimed in claim 1, wherein the origin of replication a) originates from bacteria.
- 30 3. A plasmid vector as claimed in claims 1 to 2, wherein the selection marker b) imparts a resistance to antibiotics.
4. A plasmid vector according to claims 1 to 3, wherein the promotor of element c) is selected from the group consisting of  
35 the GPD-1-, PX6-, TEF-, CUP1-, PGK-, GAP1-, TPI, PHO5-, AOX1, GAL10/CYC-1, CYC1, OliC-, ADH-, TDH-, Kex2-, MFa- and the NMT-promotor.
5. A plasmid vector according to claims 1 to 4, wherein the terminator of element c) is selected from the group consisting  
40 of the AOX1-, nos-, PGK-, TrpC- and the CYC1-terminator.
6. A plasmid vector according to claims 1 to 5, wherein the promotor of element c) is the GPD-1-promotor and the terminator  
45 of element c) is the nos-terminator.



7. A plasmid vector according to claims 1 to 6, wherein the nucleic acid sequence d) is functionally linked to a promotor facilitating recombinant expression in filamentous fungi.
- 5 8. A plasmid vector according to claims 1 to 7, wherein the nucleic acid sequence d) is functionally linked to a transcription terminator facilitating recombinant expression in filamentous fungi.
- 10 9. A selection marker comprising a nucleic acid sequence encoding a polyketide synthetase fragment, wherein said nucleic acid sequence comprises
- 15 i. a nucleic acid sequence shown in SEQ ID NO:1 or SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4 or SEQ ID NO:5; or
- ii. a nucleic acid sequence which, owing to the degeneracy of the genetic code, can be deduced from the amino acid sequence shown in SEQ ID NO:6 by back translation; or
- 20 iii. a functional equivalent of the nucleic acid sequence set forth in i) which is encoded by an amino acid sequence that has at least an identity of 50% with the SEQ ID NO: or from a functional equivalent of an amino acid sequence shown in SEQ ID NO:6 that has at least an identity of 41% with the SEQ ID NO:6 or from a functional equivalent of an amino acid sequence shown in SEQ ID NO:8 that has at least an identity of 49% with the SEQ ID NO:8 or from a functional equivalent of an amino acid sequence shown in
- 25 SEQ ID NO:10 that has at least an identity of 6; or
- 30 iv. parts of the nucleic acid sequence as defined in i., ii. or iii. consisting of at least 300bp; or
- 35 v. parts of the nucleic acid sequence as defined in i., ii. or iii. consisting of at least 300bp comprising
- a) a nucleic acid sequence shown in SEQ ID NO:7 ; or
- 40 b) a nucleic acid sequence which, owing to the degeneracy of the genetic code, can be deduced from the amino acid sequence shown in SEQ ID NO:8 by back translation; or

## 40

- c) a functional equivalent of a nucleic acid sequence set forth in a), which is encoded by amino acid sequence that has at least an identity of 85% with the SEQ ID NO:8.

5

10. Use of a nucleic acid sequence comprising

- a) a nucleic acid sequence encoding a polyketide synthetase;  
or

10

- b) parts of the nucleic acid sequence as defined in i. consisting of at least 300bp.

as marker for targeted transformation in filamentous fungi.

15

11. Use of a nucleic acid sequence according to claim 10 said nucleic acid sequence comprising

- i. a nucleic acid sequence according to claim 9; or

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- ii. a nucleic acid sequence shown in SEQ ID NO:9 or SEQ ID NO: 11; or

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- iii. a nucleic acid sequence which, owing to the degeneracy of the genetic code, can be deduced from the amino acid sequence shown in SEQ ID NO:10, SEQ ID NO:12 or SEQ ID NO:13 by back translation; or

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- iv. a functional equivalent of the nucleic acid sequence set forth in i), which is encoded by an amino acid sequence that has at least an identity of 40% with the SEQ ID NO:6 or from a functional equivalent of an amino acid sequence shown in SEQ ID NO:6 that has at least an identity of 38% with the SEQ ID NO:6 or from a functional equivalent of an amino acid sequence shown in SEQ ID NO:8 that has at least an identity of 39% with the SEQ ID NO:8 or from a functional equivalent of an amino acid sequence shown in SEQ ID NO:10 that has at least an identity of ; or

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- v. parts of the nucleic acid sequence as defined in ii., iii. or iv. consisting of at least 300bp; or

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- vi. parts of the nucleic acid sequence as defined in ii., iii or iv. consisting of at least 300bp comprising a nucleic acid sequence, which is encoded by an amino acid

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sequence that has at least an identity of 68% with the SEQ ID NO:8.

12. A plasmid vector for targeted transformation of filamentous  
5 fungi additionally comprising a selection marker comprising a  
nucleic acid sequence encoding a polyketide synthetase frag-  
ment, said nucleic acid sequence comprising
- i. a nucleic acid sequence according to claim 9; or
- 10 ii. a functional equivalent of the nucleic acid sequence set  
forth in i) which is encoded by an amino acid sequence  
that has at least an identity of 40% with the SEQ ID  
NO:6.
- 15 iii. a nucleic acid sequence shown in SEQ ID NO:9 or SEQ ID  
NO: 11;
- iv. a nucleic acid sequence which, owing to the degeneracy of  
20 the genetic code, can be deduced from the amino acid se-  
quence shown in SEQ ID NO:10, SEQ ID NO:12 or SEQ ID  
NO:13 by back translation; or
- v. parts of the nucleic acid sequence as defined in ii.,  
25 iii. or iv. consisting of at least 300bp; or
- vi. parts of the nucleic acid sequence as defined in i., ii.  
or iii. or iv. consisting of at least 300bp, which are  
30 encoded by an amino acid sequence that has at least an  
identity of 68% with SEQ ID NO:8.
13. A plasmid vector for targeted transformation of filamentous  
fungi as claimed in claims 1 to 8, additionally comprising a  
selection marker comprising a nucleic acid sequence encoding  
35 a polyketide synthetase fragment, said nucleic acid sequence  
comprising
- i. a nucleic acid sequence according to claim 9; or
- ii. a functional equivalent of the nucleic acid sequence set  
40 forth in i), which is encoded by an amino acid sequence  
that has at least an identity of 40% with the SEQ ID  
NO:6; or
- iii. a nucleic acid sequence shown in SEQ ID NO:9 or SEQ ID  
45 NO: 11;

- iv. a nucleic acid sequence which, owing to the degeneracy of the genetic code, can be deduced from the amino acid sequence shown in SEQ ID NO:10, SEQ ID NO:12 or SEQ ID NO:13 by back translation; or
- 5 v. parts of the nucleic acid sequence as defined in ii., iii. or iv. consisting of at least 300bp; or
- 10 vi. parts of the nucleic acid sequence as defined in i., ii. or iii. or iv. consisting of at least 300bp comprising a nucleic acid sequence, which is encoded by a functional equivalent of an amino acid sequence that has at least an identity of 68% with the SEQ ID NO:8.
- 15 14. An expression cassette comprising
- a) a promotor sequence in functional linkage with a nucleic acid sequence according to claim 9 in antisense orientation; and optionally
- 20 b) further genetic control sequences functionally linked to a nucleic acid sequence according to a).
15. A plasmid vector for targeted transformation of filamentous fungi additionally comprising an expression cassette according to claim 14.
- 25 16. A plasmid vector for targeted transformation of filamentous fungi as claimed in claims 1 to 8, additionally comprising an expression cassette according to claim 14.
- 30 17. A method for transforming filamentous fungi, comprising the following steps
- a) transferring a plasmid vector according to claim 12, 13, 15 or 16 into a filamentous fungi;
- 35 b) selecting successfully transformed filamentous fungi by the absence of color.
- 40 18. An expression cassette comprising
- a) a promotor sequence in functional linkage with a nucleic acid sequence comprising
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- i. a nucleic acid sequence shown in SEQ ID NO:3, 4 or 5;  
or
- 5 ii. a nucleic acid sequence which, owing to the degeneracy of the genetic code, can be deduced from the amino acid sequence shown in SEQ ID NO:6 by back translation; or
- 10 iii. a functional equivalent of the nucleic acid sequence set forth in i) which is encoded by an amino acid sequence shown in SEQ ID NO:6 that has at least an identity of 40% with the SEQ ID NO:6; or
- 15 iv. a nucleic acid sequence shown in SEQ ID NO:9 or SEQ ID NO: 11;
- 20 v. a nucleic acid sequence which, owing to the degeneracy of the genetic code, can be deduced from the amino acid sequence shown in SEQ ID NO:10, SEQ ID NO:12 or SEQ ID NO:13 by back translation;

and optionally

- 25 b) further genetic control sequences functionally linked to a nucleic acid sequence according to a).

19. A method for transformation of filamentous fungi, comprising the following steps
- 30 a) providing a filamentous fungi, in which the polyketide synthetase gene is modified in such away that the polyketide synthetase cannot be functionally expressed;
- 35 b) transforming the filamentous fungi of step a) with an expression cassette according to claim 18 or a vector comprising the aforementioned expression cassette;
- 40 c) selecting successfully transformed filamentous fungi by the presence of color.

20. A method as claimed in claim 17 or 19, wherein the plasmid vector comprises at least an additional selection marker.

21. A method as claimed in claims 17, 19 or 20, wherein the selection is confirmed by PCR.
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22. A method as claimed in claims 17, 19, 20 or 21; wherein the filamentous fungi are succesuflly transformed and identified in a high-throughput screening.

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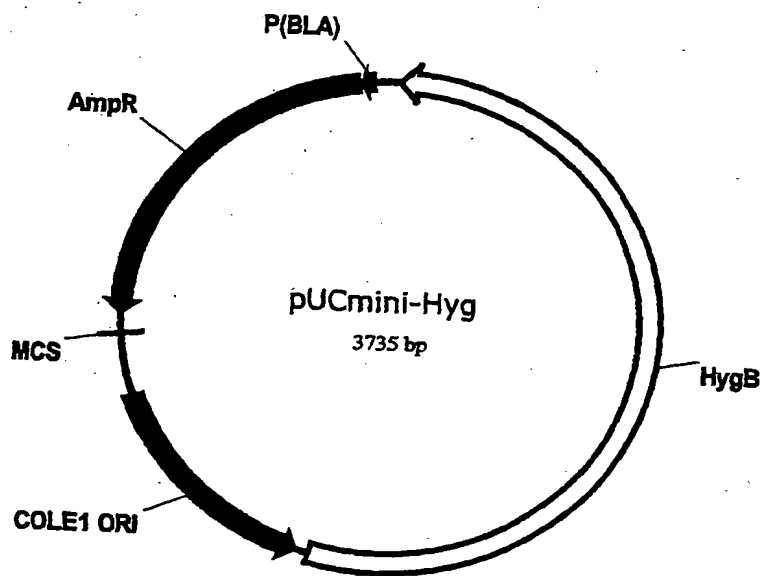


Figure 1

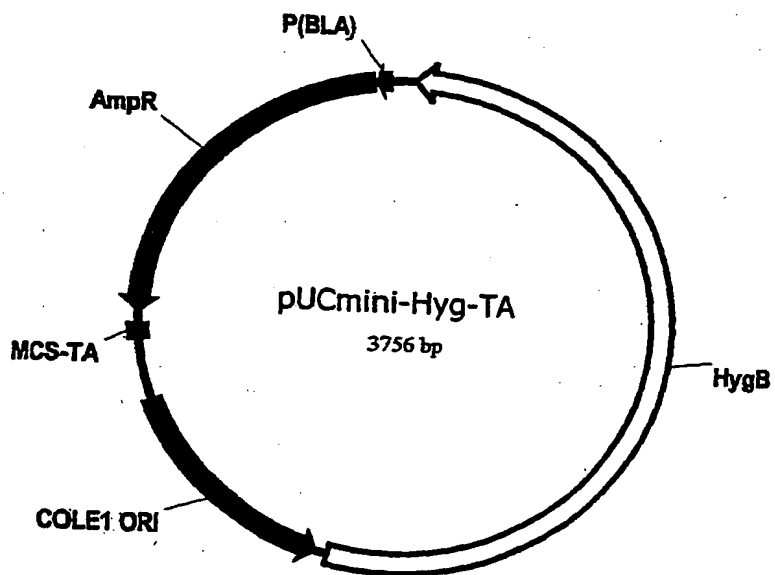


Figure 2





## SEQUENCE LISTING

&lt;110&gt; BASF Aktiengesellschaft

&lt;120&gt; Plasmid vectors for transformation of filamentous fungi

&lt;130&gt; Polyketide Synthetase

&lt;140&gt;

&lt;141&gt;

&lt;160&gt; 49

&lt;170&gt; PatentIn Ver. 2.1

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&lt;212&gt; DNA

&lt;213&gt; Fusarium graminearum

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## 10

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&lt;211&gt; 6219

&lt;212&gt; DNA

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&lt;220&gt;

&lt;221&gt; CDS

&lt;222&gt; (1)..(6216)

&lt;400&gt; 5

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cgc ttt gcc cct cca ctg aaa gac cta ctc ctc aaa ggc aac agt cct 96
Arg Phe Ala Pro Pro Leu Lys Asp Leu Leu Leu Lys Gly Asn Ser Pro
      20              25              30

tac ttg aca cat ttt gtt aaa caa gtt cac gca ctt ctt aga agg gag 144
Tyr Leu Thr His Phe Val Lys Gln Val His Ala Leu Leu Arg Arg Glu
      35              40              45

ata tca tcc ttg ccg gca gtt caa cag aag ctt ttc cca aac ttt gcc 192
Ile Ser Ser Leu Pro Ala Val Gln Gln Lys Leu Phe Pro Asn Phe Ala
      50              55              60

gac att cag gaa ctc gtc tcc aag tca gat tgg ggc agt ggt aac cct 240
Asp Ile Gln Glu Leu Val Ser Lys Ser Asp Trp Gly Ser Gly Asn Pro
      65              70              75              80

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Ala Leu Thr Ser Ala Leu Ala Cys Phe Tyr His Leu Cys Ser Phe Ile
      85              90              95

cac ttt tac gat gga caa ggt cgt acc ttt cct tcg gag aac agt cgc 336
His Phe Tyr Asp Gly Gln Gly Arg Thr Phe Pro Ser Glu Asn Ser Arg
      100              105              110

att att gga ctt tgc gtt ggt tca ctc gct gct act gct gtc agt tgc 384
Ile Ile Gly Leu Cys Val Gly Ser Leu Ala Ala Thr Ala Val Ser Cys
      115              120              125

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11

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Val Ala Leu His Val Gly Leu Arg Val Trp Arg Thr Thr Ser Leu Phe	
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Asp Val Pro Asp Arg Pro Ser Ala Thr Trp Phe Ile Ile Val Pro Glu	
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Ala Val Leu Pro Arg Glu Ser Ala Gln Asp Arg Leu Asp Ser Phe Ile	
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Ala His His Asn Met Thr Ile Ser Gly Pro Pro Ser Val Leu Glu Lys	
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Phe Ile His Ser Ile Ser Thr Ser Pro Lys Asp Ser Leu Pro Val Pro	
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Ile Tyr Ala Pro Tyr His Ala Ser His Leu Tyr Ser Met Asp Asp Val	
245 250 255	
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Asp Glu Val Leu Ser Leu Ser Ala Pro Ser Phe Ala Ser Glu Ser Ile	
260 265 270	
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Ile Pro Leu Ile Ser Ser Ser Ser Gly Asp Glu Leu Gln Pro Leu Lys	
275 280 285	
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Tyr Ala Asp Leu Leu Arg Cys Cys Val Ser Asp Met Leu Ile Gln Pro	
290 295 300	
ctg gat ctt acc aag gtc tca caa gca gtg gcc cag ctt ctc gag gtt	960
Leu Asp Leu Thr Lys Val Ser Gln Ala Val Ala Gln Leu Leu Glu Val	
305 310 315 320	
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Ser Ser Ser Thr Arg Ala Ile Ile Lys Pro Ile Ala Thr Ser Val Ser	
325 330 335	
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Asn Ser Leu Val Ser Val Leu Glu Pro Thr Leu Ala Glu Arg Cys Ala	
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## 12

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Val Asp Asn Ser Met Gly Pro Lys Ala Ser Thr Ser His Ser Ser Ala	
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gag aca caa acc gag tca tca agc aag aac tcc aaa att gcg att gtt	1152
Glu Thr Gln Thr Glu Ser Ser Ser Lys Asn Ser Lys Ile Ala Ile Val	
370 375 380	
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Ala Met Ser Gly Arg Phe Pro Asp Ala Ala Asp Leu Ser Glu Phe Trp	
385 390 395 400	
gat ctt ctc tac gaa ggt cgc gat gtt cat cga caa att ccc gag gac	1248
Asp Leu Leu Tyr Glu Gly Arg Asp Val His Arg Gln Ile Pro Glu Asp	
405 410 415	
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Arg Phe Asn Ala Glu Leu His Tyr Asp Ala Thr Gly Arg Arg Lys Asn	
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Thr Ser Lys Val Met Asn Gly Cys Phe Ile Lys Glu Pro Gly Leu Phe	
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Gly Val Phe Tyr Gly Met Thr Ser Asp Asp Trp Arg Glu Val Asn Ser	
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Gly Gln Asn Val Asp Thr Tyr Phe Ile Pro Gly Gly Asn Arg Ala Phe	
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Thr Pro Gly Arg Leu Asn Tyr Phe Phe Lys Phe Ser Gly Pro Ser Ala	
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Ser Val Asp Thr Ala Cys Ser Ser Ser Leu Val Gly Leu His Leu Ala	
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Cys Asn Ser Leu Trp Arg Asn Asp Cys Asp Thr Ala Ile Ala Gly Gly	
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Asp Gly Tyr Cys Arg Ala Asp Gly Val Gly Thr Ile Ile Leu Lys Arg	
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Leu Glu Asp Ala Glu Ala Asp Asn Asp Pro Ile Leu Gly Val Ile Leu	
625 630 635 640	
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Gly Ala Tyr Thr Asn His Ser Ala Glu Ala Val Ser Ile Thr Arg Pro	
645 650 655	
cat gcc gga gct caa gag tac atc ttc tcc aaa ctc ctc cgt gag tcg	2016
His Ala Gly Ala Gln Glu Tyr Ile Phe Ser Lys Leu Leu Arg Glu Ser	
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ggc acc gat ccc tac aac gtt agc tac atc gag atg cac ggc aca ggc	2064
Gly Thr Asp Pro Tyr Asn Val Ser Tyr Ile Glu Met His Gly Thr Gly	
675 680 685	
act caa gcc ggc gac gca acc gag atg aca tcc gtc ctc aag acg ttt	2112
Thr Gln Ala Gly Asp Ala Thr Glu Met Thr Ser Val Leu Lys Thr Phe	
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Ala Pro Thr Ser Gly Phe Gly Gly Arg Leu Pro His Gln Asn Leu His	
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Leu Gly Ser Val Lys Ala Asn Val Gly His Gly Glu Ser Ala Ser Gly	
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Ile Ile Ala Leu Ile Lys Thr Leu Leu Met Met Glu Lys Asn Met Ile	
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ccg ccg cat tgt ggt atc aag aca aag atc aat cac cat ttt cct acg	2304
Pro Pro His Cys Gly Ile Lys Thr Lys Ile Asn His His Phe Pro Thr	
755 760 765	
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Asp Leu Thr Gln Arg Asn Val His Ile Ala Lys Val Pro Thr Ser Trp	
770 775 780	
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Thr Arg Ser Gly Gln Ala Asn Pro Arg Ile Ala Phe Val Asn Asn Phe	
785 790 795 800	

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Pro Ser Val Val Ser Asp Val Thr Asp Pro Arg Thr Ser His Val Val	
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Thr Met Ser Ala Arg Ser Ala Asp Ser Leu Arg Lys Asn Leu Ala Asn	
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Asp Ser Ala Ile Glu Arg Gln Asp Val Lys Arg Ile Pro Ala Ala Ala	
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Pro Ser Val Gly Phe Val Phe Ser Gly Gln Gly Ala Gln Tyr Arg Gly	
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Met Gly Lys Glu Tyr Phe Thr Ser Phe Thr Ala Phe Arg Ser Glu Ile	
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Met Ser Tyr Asp Ser Ile Ala Gln Ala Gln Gly Phe Pro Ser Ile Leu	
945 950 955 960	
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Lys Ser Phe Gly Val Glu Pro Gly Phe Val Leu Gly His Ser Leu Gly	
995 1000 1005	
cac tat gct gct tta cac gtc gct ggt gtt ctg tcc gcc aat gat acc	3072
His Tyr Ala Ala Leu His Val Ala Gly Val Leu Ser Ala Asn Asp Thr	
1010 1015 1020	

## 15

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Gln Gln Phe Leu Asp Ala Asn Ile His Glu Val Ala Cys Val Asn Gly	
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Ser Arg Glu Val Val Ile Ser Gly Arg Val Ala Asp Ile Asp Gln Leu	
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Val Gly Leu Leu Ser Ala Asp Asn Ile Lys Ala Thr Arg Val Lys Val	
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Pro Phe Ala Phe His Ser Ala Gln Val Asp Pro Ile Leu Ser Asp Leu	
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Leu Cys Ala Leu Asp Ser Ser Val Ile Ser Pro Gly Asn His Gly Val	
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Ser Thr Leu Trp Ile Glu Ile Gly Pro His Val Val Cys Ser Thr Phe	
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Leu Lys Ser Ser Leu Gly Pro Ser Thr Pro Ala Ile Ala Ser Leu Arg	
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Arg Asn Asp Asp Cys Trp Lys Val Leu Ala Asp Gly Leu Ser Ser Leu	
1220                      1225                      1230	
tac agc agc ggg ttg aca att gac ttg aac gag tat cat cgc gac ttc	3744
Tyr Ser Ser Gly Leu Thr Ile Asp Leu Asn Glu Tyr His Arg Asp Phe	
1235                      1240                      1245	

## 16

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Ser Thr Pro Ser Val Gln Lys Ile Leu Gln Glu Thr Ser Leu Asp Gln	
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Val Leu Thr Ile Val Ala Glu Thr Asp Leu Ala Ser Pro Leu Leu Ser	
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Asn Tyr Arg Thr Asp Leu Glu Gly Tyr Ala Val Asp Val His Gly Ile	
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Glu Val His Lys Pro Leu Leu Leu Lys Glu Asp Met Asn Gly Thr Pro	
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Gln Ala Thr Pro Phe Arg Ile Glu Val Arg Tyr Pro Ile Gln Ser Thr	
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Lys His Ala Asn Cys Glu Leu Arg Leu Glu His Pro Ser Gln Trp Glu	
1425 1430 1435 1440	
gcg gag tgg gat cgc caa gcc tac ctc atc aat cgc agc gtc aac tgc	4368
Ala Glu Trp Asp Arg Gln Ala Tyr Leu Ile Asn Arg Ser Val Asn Cys	
1445 1450 1455	
ctt ctg cag cga tca gca caa ggt ttg gac agc atg ttg gca acc gga	4416
Leu Leu Gln Arg Ser Ala Gln Gly Leu Asp Ser Met Leu Ala Thr Gly	
1460 1465 1470	



## 17

atg gtc tac aag gtc ttc tcc tcc ctc gtc gac tat gcc gat ggc tac	4464
Met Val Tyr Lys Val Phe Ser Ser Leu Val Asp Tyr Ala Asp Gly Tyr	
1475 1480 1485	
aag ggt ctg cag gag gtt gtc ttg cac agc caa gag ctc gag ggc aca	4512
Lys Gly Leu Gln Glu Val Val Leu His Ser Gln Glu Leu Glu Gly Thr	
1490 1495 1500	
gca aaa gtg cgc ttc caa act ccc tcg gga ggt ttc gtc tgc aat ccc	4560
Ala Lys Val Arg Phe Gln Thr Pro Ser Gly Gly Phe Val Cys Asn Pro	
1505 1510 1515 1520	
atg tgg att gac agc tgt ggt cag acg acc ggc ttc atg atg aac tgt	4608
Met Trp Ile Asp Ser Cys Gly Gln Thr Thr Gly Phe Met Met Asn Cys	
1525 1530 1535	
cat cag act acg ccc aat gac tac gtc tac gtc aat cat ggc tgg aag	4656
His Gln Thr Thr Pro Asn Asp Tyr Val Tyr Val Asn His Gly Trp Lys	
1540 1545 1550	
tcg atg aga ttg gcc aag gcg ttc cgt gaa gat ggt acc tat cgg act	4704
Ser Met Arg Leu Ala Lys Ala Phe Arg Glu Asp Gly Thr Tyr Arg Thr	
1555 1560 1565	
tat atc cgg atg agg ccc att gat agc acc aag ttc gct ggt gac ttg	4752
Tyr Ile Arg Met Arg Pro Ile Asp Ser Thr Lys Phe Ala Gly Asp Leu	
1570 1575 1580	
tac att ctt gat gag gat gac act gtg gtt ggt gtt tat gga gac ata	4800
Tyr Ile Leu Asp Glu Asp Asp Thr Val Val Gly Val Tyr Gly Asp Ile	
1585 1590 1595 1600	
aca ttc caa ggt ttg ccg cga cga gtt ctc aac aca gtc ttg cca tct	4848
Thr Phe Gln Gly Leu Pro Arg Arg Val Leu Asn Thr Val Leu Pro Ser	
1605 1610 1615	
gcc aac gcg gtt cca gtt gat gct ccc atg ggt cga cgg gac gtg cct	4896
Ala Asn Ala Val Pro Val Asp Ala Pro Met Gly Arg Arg Asp Val Pro	
1620 1625 1630	
cca tca aga atg gat gtg cct ccc gtc agg tcc ggt gaa ggg cca ccc	4944
Pro Ser Arg Met Asp Val Pro Pro Val Arg Ser Gly Glu Gly Pro Pro	
1635 1640 1645	
act tca gca ccc acg cag caa gct atc gct ctg ccg ttc gca gcc gat	4992
Thr Ser Ala Pro Thr Gln Gln Ala Ile Ala Leu Pro Phe Ala Ala Asp	
1650 1655 1660	
aca tcc atg gac tcc cga ttg aga cct ctt ctt cgc atc ttg tca gaa	5040
Thr Ser Met Asp Ser Arg Leu Arg Pro Leu Leu Arg Ile Leu Ser Glu	
1665 1670 1675 1680	
gag atc ggt ctc ggt ctt gac gtt ctt tcg gac gat gaa ctc gac ttt	5088
Glu Ile Gly Leu Gly Leu Asp Val Leu Ser Asp Asp Glu Leu Asp Phe	
1685 1690 1695	

## 18

gcg gac cac ggt gtc gac tca ctc ctc tca ttg acc atc act ggt cgc	5136
Ala Asp His Gly Val Asp Ser Leu Leu Ser Leu Thr Ile Thr Gly Arg	
1700 1705 1710	
atg cgt gag gaa ttg ggt ctc gac gtt gaa tct aca gca ttc atg aac	5184
Met Arg Glu Glu Leu Gly Leu Asp Val Glu Ser Thr Ala Phe Met Asn	
1715 1720 1725	
tgt ccc act ttg ggc agc ttt aaa ttg ttc cta gga ctt gtc gat cag	5232
Cys Pro Thr Leu Gly Ser Phe Lys Leu Phe Leu Gly Leu Val Asp Gln	
1730 1735 1740	
gac aat aag ggc agc agc ggc agt gat ggc agt ggt agg agc agt cca	5280
Asp Asn Lys Gly Ser Ser Gly Ser Asp Gly Ser Gly Arg Ser Ser Pro	
1745 1750 1755 1760	
gca ccg ggt acc gag tct ggc gct act aca cca cct atg agc gaa gag	5328
Ala Pro Gly Thr Glu Ser Gly Ala Thr Thr Pro Pro Met Ser Glu Glu	
1765 1770 1775	
gac cag gac aag ata gtc agc agt cac tcg ctt cac cag ttc caa gcc	5376
Asp Gln Asp Lys Ile Val Ser Ser His Ser Leu His Gln Phe Gln Ala	
1780 1785 1790	
agt tcg acg ctt cta cag ggc agt ccc agt aaa gct cgc tcg act ttg	5424
Ser Ser Thr Leu Leu Gln Gly Ser Pro Ser Lys Ala Arg Ser Thr Leu	
1795 1800 1805	
ttc ttg cta cca gat ggc tcg gga tct gcc aca tcc tac gct tcc ctt	5472
Phe Leu Leu Pro Asp Gly Ser Gly Ser Ala Thr Ser Tyr Ala Ser Leu	
1810 1815 1820	
ccc ccg atc tct cca gac gga gat gtt gct gtc tac ggg ttg aac tgt	5520
Pro Pro Ile Ser Pro Asp Gly Asp Val Ala Val Tyr Gly Leu Asn Cys	
1825 1830 1835 1840	
cca tgg ctg aag gac tct agt tac ctc gtc gag ttt gga ctc aag ggc	5568
Pro Trp Leu Lys Asp Ser Ser Tyr Leu Val Glu Phe Gly Leu Lys Gly	
1845 1850 1855	
ttg aca gag ctc tat gtc aac gag ata ctc cgt cgc aag cca cag ggt	5616
Leu Thr Glu Leu Tyr Val Asn Glu Ile Leu Arg Arg Lys Pro Gln Gly	
1860 1865 1870	
cct tac aat ttg gga gga tgg tca gcc ggt ggc att tgc gct tat gaa	5664
Pro Tyr Asn Leu Gly Gly Trp Ser Ala Gly Gly Ile Cys Ala Tyr Glu	
1875 1880 1885	
gct gcc ctg atc ctc acc aga gca gga cac caa gtc gat cgc ctt atc	5712
Ala Ala Leu Ile Leu Thr Arg Ala Gly His Gln Val Asp Arg Leu Ile	
1890 1895 1900	
ttg att gac tct ccc aat ccc gtt ggt ctt gag aag cta cct cct cgc	5760
Leu Ile Asp Ser Pro Asn Pro Val Gly Leu Glu Lys Leu Pro Pro Arg	
1905 1910 1915 1920	

ttg tac gat ttc ctc aat tcg cag aat gtc ttt gga tca gac aac ccg 5808  
 Leu Tyr Asp Phe Leu Asn Ser Gln Asn Val Phe Gly Ser Asp Asn Pro  
 1925 1930 1935

cac agc act gct gga aca agc gtc aaa gct cca gaa tgg ctt ctt gca 5856  
 His Ser Thr Ala Gly Thr Ser Val Lys Ala Pro Glu Trp Leu Leu Ala  
 1940 1945 1950

cat ttc ctg gcc ttc att gac gct ctg gat gct tat gtc gca gtg cct 5904  
 His Phe Leu Ala Phe Ile Asp Ala Leu Asp Ala Tyr Val Ala Val Pro  
 1955 1960 1965

tgg gac tct ggt cta gtc ggt cta gca tca ccg ctc cct gca ccg ccg 5952  
 Trp Asp Ser Gly Leu Val Gly Leu Ala Ser Pro Leu Pro Ala Pro Pro  
 1970 1975 1980

cag aca tac atg ctg tgg gca gaa gac gga gtt tgc aaa gac tct gat 6000  
 Gln Thr Tyr Met Leu Trp Ala Glu Asp Gly Val Cys Lys Asp Ser Asp  
 1985 1990 1995 2000

agt gct cgt ccc gag tac cgt gac gat gac cca cgc gag atg aga tgg 6048  
 Ser Ala Arg Pro Glu Tyr Arg Asp Asp Asp Pro Arg Glu Met Arg Trp  
 2005 2010 2015

ctg ttg gag aac aga aca aac ttt ggt ccg aat ggt tgg gag gcg cta 6096  
 Leu Leu Glu Asn Arg Thr Asn Phe Gly Pro Asn Gly Trp Glu Ala Leu  
 2020 2025 2030

ctt ggt ggt aaa gag ggt ttg ttc atg gat cgg att gcg gaa gcg aat 6144  
 Leu Gly Gly Lys Glu Gly Leu Phe Met Asp Arg Ile Ala Glu Ala Asn  
 2035 2040 2045

cat ttt agt atg ttg aag aga gga cgg aat gcg gaa tat gtc tct gca 6192  
 His Phe Ser Met Leu Lys Arg Gly Arg Asn Ala Glu Tyr Val Ser Ala  
 2050 2055 2060

ttc ctg gct cgg gcc ttg gac aat tag 6219  
 Phe Leu Ala Arg Ala Leu Asp Asn  
 2065 2070

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 <212> PRT  
 <213> *Fusarium graminearum*

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 Arg Phe Ala Pro Pro Leu Lys Asp Leu Leu Leu Lys Gly Asn Ser Pro  
 20 25 30  
 Tyr Leu Thr His Phe Val Lys Gln Val His Ala Leu Leu Arg Arg Glu  
 35 40 45

## 20

Ile Ser Ser Leu Pro Ala Val Gln Gln Lys Leu Phe Pro Asn Phe Ala  
 50 55 60

Asp Ile Gln Glu Leu Val Ser Lys Ser Asp Trp Gly Ser Gly Asn Pro  
 65 70 75 80

Ala Leu Thr Ser Ala Leu Ala Cys Phe Tyr His Leu Cys Ser Phe Ile  
 85 90 95

His Phe Tyr Asp Gly Gln Gly Arg Thr Phe Pro Ser Glu Asn Ser Arg  
 100 105 110

Ile Ile Gly Leu Cys Val Gly Ser Leu Ala Ala Thr Ala Val Ser Cys  
 115 120 125

Ser Thr Ser Leu Ser Glu Leu Val Ser Ala Gly Val Asp Ala Val Arg  
 130 135 140

Val Ala Leu His Val Gly Leu Arg Val Trp Arg Thr Thr Ser Leu Phe  
 145 150 155 160

Asp Val Pro Asp Arg Pro Ser Ala Thr Trp Phe Ile Ile Val Pro Glu  
 165 170 175

Ala Val Leu Pro Arg Glu Ser Ala Gln Asp Arg Leu Asp Ser Phe Ile  
 180 185 190

Ile Glu Met Gly Leu Ala Arg Ser Ser Val Pro Tyr Ile Ser Ser Val  
 195 200 205

Ala His His Asn Met Thr Ile Ser Gly Pro Pro Ser Val Leu Glu Lys  
 210 215 220

Phe Ile His Ser Ile Ser Thr Ser Pro Lys Asp Ser Leu Pro Val Pro  
 225 230 235 240

Ile Tyr Ala Pro Tyr His Ala Ser His Leu Tyr Ser Met Asp Asp Val  
 245 250 255

Asp Glu Val Leu Ser Leu Ser Ala Pro Ser Phe Ala Ser Glu Ser Ile  
 260 265 270

Ile Pro Leu Ile Ser Ser Ser Ser Gly Asp Glu Leu Gln Pro Leu Lys  
 275 280 285

Tyr Ala Asp Leu Leu Arg Cys Cys Val Ser Asp Met Leu Ile Gln Pro  
 290 295 300

Leu Asp Leu Thr Lys Val Ser Gln Ala Val Ala Gln Leu Leu Glu Val  
 305 310 315 320

Ser Ser Ser Thr Arg Ala Ile Ile Lys Pro Ile Ala Thr Ser Val Ser  
 325 330 335

Asn Ser Leu Val Ser Val Leu Glu Pro Thr Leu Ala Glu Arg Cys Ala  
 340 345 350

## 21

Val	Asp	Asn	Ser	Met	Gly	Pro	Lys	Ala	Ser	Thr	Ser	His	Ser	Ser	Ala	355	360	365	
Glu	Thr	Gln	Thr	Glu	Ser	Ser	Ser	Lys	Asn	Ser	Lys	Ile	Ala	Ile	Val	370	375	380	
Ala	Met	Ser	Gly	Arg	Phe	Pro	Asp	Ala	Ala	Asp	Leu	Ser	Glu	Phe	Trp	385	390	395	400
Asp	Leu	Leu	Tyr	Glu	Gly	Arg	Asp	Val	His	Arg	Gln	Ile	Pro	Glu	Asp	405	410	415	
Arg	Phe	Asn	Ala	Glu	Leu	His	Tyr	Asp	Ala	Thr	Gly	Arg	Arg	Lys	Asn	420	425	430	
Thr	Ser	Lys	Val	Met	Asn	Gly	Cys	Phe	Ile	Lys	Glu	Pro	Gly	Leu	Phe	435	440	445	
Asp	Ala	Arg	Phe	Phe	Asn	Met	Ser	Pro	Lys	Glu	Ala	Glu	Gln	Ser	Asp	450	455	460	
Pro	Gly	Gln	Arg	Met	Ala	Leu	Glu	Thr	Ala	Tyr	Glu	Ala	Leu	Glu	Met	465	470	475	480
Ala	Ser	Ile	Val	Pro	Asp	Arg	Thr	Pro	Ser	Thr	Gln	Arg	Asp	Arg	Val	485	490	495	
Gly	Val	Phe	Tyr	Gly	Met	Thr	Ser	Asp	Asp	Trp	Arg	Glu	Val	Asn	Ser	500	505	510	
Gly	Gln	Asn	Val	Asp	Thr	Tyr	Phe	Ile	Pro	Gly	Gly	Asn	Arg	Ala	Phe	515	520	525	
Thr	Pro	Gly	Arg	Leu	Asn	Tyr	Phe	Phe	Lys	Phe	Ser	Gly	Pro	Ser	Ala	530	535	540	
Ser	Val	Asp	Thr	Ala	Cys	Ser	Ser	Ser	Leu	Val	Gly	Leu	His	Leu	Ala	545	550	555	560
Cys	Asn	Ser	Leu	Trp	Arg	Asn	Asp	Cys	Asp	Thr	Ala	Ile	Ala	Gly	Gly	565	570	575	
Thr	Asn	Val	Met	Thr	Asn	Pro	Asp	Asn	Phe	Ala	Gly	Leu	Asp	Arg	Gly	580	585	590	
His	Phe	Leu	Ser	Arg	Thr	Gly	Asn	Cys	Asn	Thr	Phe	Asp	Asp	Gly	Ala	595	600	605	
Asp	Gly	Tyr	Cys	Arg	Ala	Asp	Gly	Val	Gly	Thr	Ile	Ile	Leu	Lys	Arg	610	615	620	
Leu	Glu	Asp	Ala	Glu	Ala	Asp	Asn	Asp	Pro	Ile	Leu	Gly	Val	Ile	Leu	625	630	635	640
Gly	Ala	Tyr	Thr	Asn	His	Ser	Ala	Glu	Ala	Val	Ser	Ile	Thr	Arg	Pro	645	650	655	

His Ala Gly Ala Gln Glu Tyr Ile Phe Ser Lys Leu Leu Arg Glu Ser  
 660 665 670  
 Gly Thr Asp Pro Tyr Asn Val Ser Tyr Ile Glu Met His Gly Thr Gly  
 675 680 685  
 Thr Gln Ala Gly Asp Ala Thr Glu Met Thr Ser Val Leu Lys Thr Phe  
 690 695 700  
 Ala Pro Thr Ser Gly Phe Gly Gly Arg Leu Pro His Gln Asn Leu His  
 705 710 715 720  
 Leu Gly Ser Val Lys Ala Asn Val Gly His Gly Glu Ser Ala Ser Gly  
 725 730 735  
 Ile Ile Ala Leu Ile Lys Thr Leu Leu Met Met Glu Lys Asn Met Ile  
 740 745 750  
 Pro Pro His Cys Gly Ile Lys Thr Lys Ile Asn His His Phe Pro Thr  
 755 760 765  
 Asp Leu Thr Gln Arg Asn Val His Ile Ala Lys Val Pro Thr Ser Trp  
 770 775 780  
 Thr Arg Ser Gly Gln Ala Asn Pro Arg Ile Ala Phe Val Asn Asn Phe  
 785 790 795 800  
 Ser Ala Ala Gly Gly Asn Ser Ala Val Leu Leu Gln Asp Ala Pro Gln  
 805 810 815  
 Pro Ser Val Val Ser Asp Val Thr Asp Pro Arg Thr Ser His Val Val  
 820 825 830  
 Thr Met Ser Ala Arg Ser Ala Asp Ser Leu Arg Lys Asn Leu Ala Asn  
 835 840 845  
 Leu Lys Glu Leu Val Glu Gly Gln Gly Asp Ser Glu Val Gly Phe Leu  
 850 855 860  
 Ser Lys Leu Ser Tyr Thr Thr Thr Ala Arg Arg Met His His Gln Phe  
 865 870 875 880  
 Arg Ala Ser Val Thr Ala Gln Thr Arg Glu Gln Leu Leu Lys Gly Leu  
 885 890 895  
 Asp Ser Ala Ile Glu Arg Gln Asp Val Lys Arg Ile Pro Ala Ala Ala  
 900 905 910  
 Pro Ser Val Gly Phe Val Phe Ser Gly Gln Gly Ala Gln Tyr Arg Gly  
 915 920 925  
 Met Gly Lys Glu Tyr Phe Thr Ser Phe Thr Ala Phe Arg Ser Glu Ile  
 930 935 940  
 Met Ser Tyr Asp Ser Ile Ala Gln Ala Gln Gly Phe Pro Ser Ile Leu  
 945 950 955 960

## 23

Pro Leu Ile Arg Gly Glu Val Glu Ala Asp Ser Leu Ser Pro Val Glu  
 965 970 975  
 Ile Gln Leu Gly Leu Thr Cys Leu Gln Met Ala Leu Ala Lys Leu Trp  
 980 985 990  
 Lys Ser Phe Gly Val Glu Pro Gly Phe Val Leu Gly His Ser Leu Gly  
 995 1000 1005  
 His Tyr Ala Ala Leu His Val Ala Gly Val Leu Ser Ala Asn Asp Thr  
 1010 1015 1020  
 Ile Tyr Leu Thr Gly Ile Arg Ala Gln Leu Leu Val Asp Lys Cys Gln  
 1025 1030 1035 1040  
 Ala Gly Thr His Ser Met Leu Ala Val Arg Ala Ser Leu Leu Gln Ile  
 1045 1050 1055  
 Gln Gln Phe Leu Asp Ala Asn Ile His Glu Val Ala Cys Val Asn Gly  
 1060 1065 1070  
 Ser Arg Glu Val Val Ile Ser Gly Arg Val Ala Asp Ile Asp Gln Leu  
 1075 1080 1085  
 Val Gly Leu Leu Ser Ala Asp Asn Ile Lys Ala Thr Arg Val Lys Val  
 1090 1095 1100  
 Pro Phe Ala Phe His Ser Ala Gln Val Asp Pro Ile Leu Ser Asp Leu  
 1105 1110 1115 1120  
 Asp Thr Ala Ala Ser Arg Val Thr Phe His Ser Leu Gln Ile Pro Val  
 1125 1130 1135  
 Leu Cys Ala Leu Asp Ser Ser Val Ile Ser Pro Gly Asn His Gly Val  
 1140 1145 1150  
 Ile Gly Pro Leu His Leu Gln Arg His Cys Arg Glu Thr Val Asn Phe  
 1155 1160 1165  
 Glu Gly Ala Leu His Ala Ala Glu His Glu Lys Ile Ile Asn Lys Thr  
 1170 1175 1180  
 Ser Thr Leu Trp Ile Glu Ile Gly Pro His Val Val Cys Ser Thr Phe  
 1185 1190 1195 1200  
 Leu Lys Ser Ser Leu Gly Pro Ser Thr Pro Ala Ile Ala Ser Leu Arg  
 1205 1210 1215  
 Arg Asn Asp Asp Cys Trp Lys Val Leu Ala Asp Gly Leu Ser Ser Leu  
 1220 1225 1230  
 Tyr Ser Ser Gly Leu Thr Ile Asp Leu Asn Glu Tyr His Arg Asp Phe  
 1235 1240 1245  
 Lys Ala Ser His Gln Val Leu Arg Leu Pro Cys Tyr Ser Trp Glu His  
 1250 1255 1260

## 24

Lys Asn Tyr Trp Ile Gln Tyr Lys Tyr Asp Trp Ser Leu Ala Lys Gly  
 265 1270 1275 1280  
 Asp Pro Pro Ile Ala Pro Asn Ser Ser Val Glu Ala Val Ser Ala Leu  
 1285 1290 1295  
 Ser Thr Pro Ser Val Gln Lys Ile Leu Gln Glu Thr Ser Leu Asp Gln  
 1300 1305 1310  
 Val Leu Thr Ile Val Ala Glu Thr Asp Leu Ala Ser Pro Leu Leu Ser  
 1315 1320 1325  
 Glu Val Ala Gln Gly His Arg Val Asn Gly Val Lys Val Cys Thr Ser  
 1330 1335 1340  
 Ser Val Tyr Ala Asp Val Gly Leu Thr Leu Gly Lys Tyr Ile Leu Asp  
 345 1350 1355 1360  
 Asn Tyr Arg Thr Asp Leu Glu Gly Tyr Ala Val Asp Val His Gly Ile  
 1365 1370 1375  
 Glu Val His Lys Pro Leu Leu Leu Lys Glu Asp Met Asn Gly Thr Pro  
 1380 1385 1390  
 Gln Ala Thr Pro Phe Arg Ile Glu Val Arg Tyr Pro Ile Gln Ser Thr  
 1395 1400 1405  
 Thr Ala Leu Met Ser Ile Ser Thr Thr Gly Pro Asn Gly Gln His Ile  
 1410 1415 1420  
 Lys His Ala Asn Cys Glu Leu Arg Leu Glu His Pro Ser Gln Trp Glu  
 425 1430 1435 1440  
 Ala Glu Trp Asp Arg Gln Ala Tyr Leu Ile Asn Arg Ser Val Asn Cys  
 1445 1450 1455  
 Leu Leu Gln Arg Ser Ala Gln Gly Leu Asp Ser Met Leu Ala Thr Gly  
 1460 1465 1470  
 Met Val Tyr Lys Val Phe Ser Ser Leu Val Asp Tyr Ala Asp Gly Tyr  
 1475 1480 1485  
 Lys Gly Leu Gln Glu Val Val Leu His Ser Gln Glu Leu Glu Gly Thr  
 1490 1495 1500  
 Ala Lys Val Arg Phe Gln Thr Pro Ser Gly Gly Phe Val Cys Asn Pro  
 505 1510 1515 1520  
 Met Trp Ile Asp Ser Cys Gly Gln Thr Thr Gly Phe Met Met Asn Cys  
 1525 1530 1535  
 His Gln Thr Thr Pro Asn Asp Tyr Val Tyr Val Asn His Gly Trp Lys  
 1540 1545 1550  
 Ser Met Arg Leu Ala Lys Ala Phe Arg Glu Asp Gly Thr Tyr Arg Thr  
 1555 1560 1565



## 25

Tyr Ile Arg Met Arg Pro Ile Asp Ser Thr Lys Phe Ala Gly Asp Leu  
 1570 1575 1580  
 Tyr Ile Leu Asp Glu Asp Asp Thr Val Val Gly Val Tyr Gly Asp Ile  
 585 1590 1595 1600  
 Thr Phe Gln Gly Leu Pro Arg Arg Val Leu Asn Thr Val Leu Pro Ser  
 1605 1610 1615  
 Ala Asn Ala Val Pro Val Asp Ala Pro Met Gly Arg Arg Asp Val Pro  
 1620 1625 1630  
 Pro Ser Arg Met Asp Val Pro Pro Val Arg Ser Gly Glu Gly Pro Pro  
 1635 1640 1645  
 Thr Ser Ala Pro Thr Gln Gln Ala Ile Ala Leu Pro Phe Ala Ala Asp  
 1650 1655 1660  
 Thr Ser Met Asp Ser Arg Leu Arg Pro Leu Leu Arg Ile Leu Ser Glu  
 665 1670 1675 1680  
 Glu Ile Gly Leu Gly Leu Asp Val Leu Ser Asp Asp Glu Leu Asp Phe  
 1685 1690 1695  
 Ala Asp His Gly Val Asp Ser Leu Leu Ser Leu Thr Ile Thr Gly Arg  
 1700 1705 1710  
 Met Arg Glu Glu Leu Gly Leu Asp Val Glu Ser Thr Ala Phe Met Asn  
 1715 1720 1725  
 Cys Pro Thr Leu Gly Ser Phe Lys Leu Phe Leu Gly Leu Val Asp Gln  
 1730 1735 1740  
 Asp Asn Lys Gly Ser Ser Gly Ser Asp Gly Ser Gly Arg Ser Ser Pro  
 745 1750 1755 1760  
 Ala Pro Gly Thr Glu Ser Gly Ala Thr Thr Pro Pro Met Ser Glu Glu  
 1765 1770 1775  
 Asp Gln Asp Lys Ile Val Ser Ser His Ser Leu His Gln Phe Gln Ala  
 1780 1785 1790  
 Ser Ser Thr Leu Leu Gln Gly Ser Pro Ser Lys Ala Arg Ser Thr Leu  
 1795 1800 1805  
 Phe Leu Leu Pro Asp Gly Ser Gly Ser Ala Thr Ser Tyr Ala Ser Leu  
 1810 1815 1820  
 Pro Pro Ile Ser Pro Asp Gly Asp Val Ala Val Tyr Gly Leu Asn Cys  
 825 1830 1835 1840  
 Pro Trp Leu Lys Asp Ser Ser Tyr Leu Val Glu Phe Gly Leu Lys Gly  
 1845 1850 1855  
 Leu Thr Glu Leu Tyr Val Asn Glu Ile Leu Arg Arg Lys Pro Gln Gly  
 1860 1865 1870

## 26

Pro Tyr Asn Leu Gly Gly Trp Ser Ala Gly Gly Ile Cys Ala Tyr Glu  
 1875 1880 1885

Ala Ala Leu Ile Leu Thr Arg Ala Gly His Gln Val Asp Arg Leu Ile  
 1890 1895 1900

Leu Ile Asp Ser Pro Asn Pro Val Gly Leu Glu Lys Leu Pro Pro Arg  
 905 1910 1915 1920

Leu Tyr Asp Phe Leu Asn Ser Gln Asn Val Phe Gly Ser Asp Asn Pro  
 1925 1930 1935

His Ser Thr Ala Gly Thr Ser Val Lys Ala Pro Glu Trp Leu Leu Ala  
 1940 1945 1950

His Phe Leu Ala Phe Ile Asp Ala Leu Asp Ala Tyr Val Ala Val Pro  
 1955 1960 1965

Trp Asp Ser Gly Leu Val Gly Leu Ala Ser Pro Leu Pro Ala Pro Pro  
 1970 1975 1980

Gln Thr Tyr Met Leu Trp Ala Glu Asp Gly Val Cys Lys Asp Ser Asp  
 985 1990 1995 2000

Ser Ala Arg Pro Glu Tyr Arg Asp Asp Asp Pro Arg Glu Met Arg Trp  
 2005 2010 2015

Leu Leu Glu Asn Arg Thr Asn Phe Gly Pro Asn Gly Trp Glu Ala Leu  
 2020 2025 2030

Leu Gly Gly Lys Glu Gly Leu Phe Met Asp Arg Ile Ala Glu Ala Asn  
 2035 2040 2045

His Phe Ser Met Leu Lys Arg Gly Arg Asn Ala Glu Tyr Val Ser Ala  
 2050 2055 2060

Phe Leu Ala Arg Ala Leu Asp Asn  
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<213> *Fusarium graminearum*

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 Gly Asn Arg Ala Phe Thr Pro Gly Arg Leu Asn Tyr Phe Phe Lys Phe  
 1 5 10 15

agt ggg cct agc gct agt gtt gat acg gct tgc tcc tcc agt ctc gtt 96  
 Ser Gly Pro Ser Ala Ser Val Asp Thr Ala Cys Ser Ser Leu Val  
 20 25 30

## 27

ggc ttg cac ttg gct tgt aat tcc ctc tgg aga aat gat tgc gat aca 144  
 Gly Leu His Leu Ala Cys Asn Ser Leu Trp Arg Asn Asp Cys Asp Thr  
           35                          40                          45

gct att gcg ggc gga acc aat gtc atg act aac cct gac aac ttc gct 192  
 Ala Ile Ala Gly Gly Thr Asn Val Met Thr Asn Pro Asp Asn Phe Ala  
           50                          55                          60

ggt ttg gac cga ggc cac ttc cta tct aga acc ggt 228  
 Gly Leu Asp Arg Gly His Phe Leu Ser Arg Thr Gly  
           65                          70                          75

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<212> PRT

<213> Fusarium graminearum

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Ser Gly Pro Ser Ala Ser Val Asp Thr Ala Cys Ser Ser Ser Leu Val  
                           20                          25                          30

Gly Leu His Leu Ala Cys Asn Ser Leu Trp Arg Asn Asp Cys Asp Thr  
           35                          40                          45

Ala Ile Ala Gly Gly Thr Asn Val Met Thr Asn Pro Asp Asn Phe Ala  
           50                          55                          60

Gly Leu Asp Arg Gly His Phe Leu Ser Arg Thr Gly  
           65                          70                          75

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<213> Wagiella dermatidis

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 gtcttttacc ggaaggacaa tgtcctcttg cagagcttcc tggaaagagc tgggtgaagcc 120  
 gtcagattcg aaaaccagaa ccatcacat ccctccaaag cagtccctaa tttctctacc 180  
 atccaagagc ttgtcgaccg ttattacaga ggtgacgcga aggatgccgc gaccgagagt 240  
 gctctcgttt gcatttcaca gttttgtcat ttcatgtgtg cctttgagga gagacggcct 300  
 tcgtatattc aacctaattc agatgctcga ctagtcggtc tgtgcacagg tctaattgct 360  
 gcgaccgcag tcgtgcctc agattccctg acagccctta ttcctttggc ggtcgaagcg 420  
 gtccggatag cgttcagagc tgggtgccac gtgggaaaag ttgcacagca gacagaatgt 480  
 gactccaaga ctcaatcgtg gtccaccatc gttgctgccg acgagaaatc tgcgcaggag 540  
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 gttttcttcg ccacttcagt caccattagc gttccgccat ggaccaaagc cgtcttttn 660  
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 cagcatcgc acntacactc gcaatccgat ttggacaaaa tcttacgacc gcagaccaag 780  
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&lt;210&gt; 10

&lt;211&gt; 2177

&lt;212&gt; PRT

&lt;213&gt; Wagiella dermatidis

&lt;400&gt; 10

Met Glu Glu Val Tyr Val Phe Gly Asp Gln Thr Ala Asp Cys Arg Ala  
 1 5 10 15

Phe Phe Thr Lys Val Phe Thr Arg Lys Asp Asn Val Leu Leu Gln Ser  
 20 25 30

Phe Leu Glu Arg Ala Gly Glu Ala Val Arg Phe Glu Asn Gln Asn Arg  
 35 40 45

Ser His Pro Ser Lys Ala Val Pro Asn Phe Ser Thr Ile Gln Glu Leu  
 50 55 60

Val Asp Arg Tyr Tyr Arg Gly Asp Ala Lys Asp Ala Ala Thr Glu Ser  
 65 70 75 80

Ala Leu Val Cys Ile Ser Gln Phe Cys His Phe Ile Gly Ala Phe Glu  
 85 90 95

Glu Arg Arg Pro Ser Tyr Ile Gln Pro Asn Ser Asp Ala Arg Leu Val  
 100 105 110

Gly Leu Cys Thr Gly Leu Ile Ala Ala Thr Ala Val Ala Ala Ser Asp  
 115 120 125

Ser Leu Thr Ala Leu Ile Pro Leu Ala Val Glu Ala Val Arg Ile Ala  
 130 135 140

Phe Arg Ala Gly Ala His Val Gly Lys Val Ala Gln Gln Thr Glu Cys  
 145 150 155 160

Asp Ser Lys Thr Gln Ser Trp Ser Thr Ile Val Ala Ala Asp Glu Lys  
 165 170 175

Ser Ala Gln Glu Ala Leu Asp Ala Phe His Lys Glu Xaa Gly Thr Ser  
 180 185 190

Pro Ile Asn Gln Leu Trp Ile Ser Val Ser Ser Ala Thr Ser Val Thr  
 195 200 205

Ile Ser Val Pro Pro Trp Thr Lys Ala Arg Leu Xaa Glu Glu Ser Glu  
 210 215 220

Phe Phe Arg Thr Gln Lys Ser Ala Pro Val Ser Ile Phe Ala Pro Tyr  
 225 230 235 240

His Ala Ser His Xaa His Ser Gln Ser Asp Leu Asp Lys Ile Leu Arg  
 245 250 255

Pro Gln Thr Lys Thr Ile Phe Gly Asn Thr Thr Val Arg Phe Pro Val  
 260 265 270

## 31

Cys Ser Ser Val Thr Gly Lys Pro Phe Asn Ala Glu Asn Gly Phe Glu  
 275 280 285  
 Leu Leu Gln Ala Ala Leu Lys Glu Ile Ile Ile Asp Pro Leu Arg Trp  
 290 295 300  
 Asp Lys Val Leu Lys Tyr Cys Ala Ala Gly Lys Ala Ser Glu Ala Lys  
 305 310 315 320  
 Val Phe Ala Val Gly Pro Thr Asn Leu Ala Ser Ser Val Val Ser Ala  
 325 330 335  
 Leu Lys Ala Ser Thr Thr Lys Val Thr Leu Glu Asp His Ser Thr Trp  
 340 345 350  
 Ser Thr Val Pro Pro Gln Gly Thr Arg His Ser Lys Arg Glu Ala Asp  
 355 360 365  
 Ile Ala Ile Val Gly Phe Ser Gly Arg Phe Pro Asp Ala Ala Asp Asn  
 370 375 380  
 Glu Leu Phe Trp Gln Leu Leu Glu Arg Gly Leu Asp Val His Arg Pro  
 385 390 395 400  
 Val Pro Pro Asp Arg Phe Pro Val Glu Ser His Thr Asp Pro Ser Gly  
 405 410 415  
 Lys Lys Lys Asn Thr Ser His Thr Pro Phe Gly Asn Phe Ile Glu Lys  
 420 425 430  
 Pro Gly Leu Phe Asp Ala Arg Phe Phe Asn Met Ser Pro Arg Glu Ala  
 435 440 445  
 Ala Gln Thr Asp Pro Met Gln Arg Leu Met Leu Thr Thr Gly Tyr Xaa  
 450 455 460  
 Ala Met Glu Met Ala Gly Ile Val Pro Gly Xaa Thr Pro Ser Thr Xaa  
 465 470 475 480  
 His Asp Arg Ile Gly Thr Phe Tyr Gly Gln Thr Ser Xaa Xaa Trp Arg  
 485 490 495  
 Glu Val Asn Ala Ala Xaa Asp Ile Asp Thr Tyr Phe Ile Ser Gly Gly  
 500 505 510  
 Val Arg Ala Phe Gly Pro Gly Xaa Ile Asn Tyr Phe Phe Lys Phe Ser  
 515 520 525  
 Gly Pro Xaa Phe Ser Val Asp Met Xaa Ala Asn Pro Ala Trp Pro Xaa  
 530 535 540  
 Met Asn Val Ala Ile Thr Ser Leu Arg Ala Asn Glu Cys Asp Thr Val  
 545 550 555 560  
 Phe Thr Gly Gly Ala Asn Val Leu Thr Asn Ser Asp Ile Phe Ser Gly  
 565 570 575

## 32

Leu Ser Arg Gly His Phe Leu Ser Lys Thr Gly Ser Cys Lys Thr Trp  
 580 585 590  
 Asp Asn Asp Ala Asp Gly Tyr Cys Arg Gly Asp Gly Val Cys Thr Val  
 595 600 605  
 Ile Met Lys Arg Leu Asp Asp Ala Leu Ala Asp Arg Asp Pro Val Leu  
 610 615 620  
 Gly Val Ile Arg Gly Ile Gly Thr Asn His Ser Ala Glu Ala Val Ser  
 625 630 635 640  
 Ile Thr His Pro Cys Ala Glu Asn Gln Ala Phe Leu Phe Asp Lys Val  
 645 650 655  
 Leu Lys Glu Cys Asn Val His Cys Asn Asp Val Asn Tyr Val Glu Met  
 660 665 670  
 His Gly Thr Gly Thr Gln Ala Gly Asp Gly Ile Glu Met Glu Ser Val  
 675 680 685  
 Ser Ser Val Phe Ala Pro Arg Gln Pro Arg Arg Arg Pro Asp Gln Pro  
 690 695 700  
 Leu Tyr Val Gly Ala Val Lys Ser Asn Ile Gly His Gly Glu Ala Val  
 705 710 715 720  
 Ser Gly Val Ser Ala Leu Ile Lys Val Leu Leu Met Leu Gln Lys Asn  
 725 730 735  
 Lys Ile Pro Pro His Thr Gly Ile Lys Lys Gln Ile Asn Lys Asn Phe  
 740 745 750  
 Ala Pro Asp Leu Lys Glu Arg Asn Val Asn Ile Ala Phe Gln Thr Thr  
 755 760 765  
 Pro Phe Pro Arg Pro Pro Gly Gly Lys Arg Thr Val Phe Ile Asn Asn  
 770 775 780  
 Phe Ser Ala Ala Gly Gly Asn Thr Ala Met Leu Leu Gln Asp Gly Pro  
 785 790 795 800  
 Glu Val Pro Thr Glu Pro Ser Ser Asp Pro Arg Ser Thr His Val Val  
 805 810 815  
 Thr Xaa Ser Ala Lys Ser Leu Ala Ala Phe Lys Arg Thr Leu Ala Lys  
 820 825 830  
 Tyr Glu Ala Tyr Leu Asn Ala His Pro Asn Val Gly Leu Pro Asp Leu  
 835 840 845  
 Ala Tyr Thr Val Thr Ala Arg Arg Ala His Tyr Ser Leu Pro Arg Arg  
 850 855 860  
 Phe Pro Val Gln Ser Ile Ser Gln Leu Gln Ala Ser Leu Arg Ala Ile  
 865 870 875 880



Gln Asp Gln Thr His Asn Pro Ile Pro Leu Ala Ser Pro Gln Ile Ala  
 885 890 895  
 Met Ala Phe Thr Gly Gln Gly Ser Gln Tyr Thr Gly Met Gly Gln Lys  
 900 905 910  
 Leu Phe Glu Thr Ser Lys Gln Phe Arg Gln Asp Ile Glu Glu Phe Asn  
 915 920 925  
 Glu Ile Ala Leu Arg Gln Gly Leu Pro Ser Ile Met Pro Leu Ile Asp  
 930 935 940  
 Gly Ser Val Glu Val Gln His Leu Pro Pro Thr Val Val Gln Leu Gly  
 945 950 955 960  
 Met Cys Cys Ile Gln Met Ala Leu Thr His Leu Trp Ser Thr Trp Gly  
 965 970 975  
 Ile Gln Pro Ser Val Val Ile Gly His Ser Leu Gly Glu Tyr Ala Ala  
 980 985 990  
 Leu Gln Ala Ala Gly Val Leu Ser Ile Ala Asp Thr Ile Tyr Leu Val  
 995 1000 1005  
 Gly Lys Arg Ala Gln Leu Leu Glu Gln Lys Cys Thr Ala Gly Thr His  
 1010 1015 1020  
 Ala Met Leu Ala Val Arg Ser Pro Val Gly Gly Leu Gln Asp Val Val  
 1025 1030 1035 1040  
 Ala Asn Ser His Gly Lys Ile Glu Asn Cys Gly Ile Asn Gly Val Ser  
 1045 1050 1055  
 Asp Thr Val Leu Ser Gly Thr Met Gly Asp Ile Asp Thr Val Ala Gln  
 1060 1065 1070  
 Lys Leu Ala Asp Ala Gly Gln Lys Cys Thr Lys Leu Lys Leu Pro Phe  
 1075 1080 1085  
 Ala Phe His Ser Ser Gln Val Asp Pro Ile Leu Ala Asp Phe Glu Lys  
 1090 1095 1100  
 Leu Ala Ser Ser Val Asn Tyr His Pro Pro Arg Val Pro Val Ile Ser  
 1105 1110 1115 1120  
 Pro Leu Leu Ser Asp Val Val Ser Val Gly Gly Val Phe Asp Ala Phe  
 1125 1130 1135  
 Tyr Leu Ser Arg His Cys Arg Lys Thr Val Asp Phe Val Gly Gly Leu  
 1140 1145 1150  
 Ser Ala Gly Met Ser Thr Ala Thr Ile Ser Asp Thr Ser Leu Trp Leu  
 1155 1160 1165  
 Glu Val Gly Gly His Pro Leu Cys Ala Ser Met Ile Lys Ser Cys Leu  
 1170 1175 1180

## 34

Ser Val Pro Thr Leu Ala Thr Met Arg Arg Asp Glu Asp Pro Trp Lys  
 1185 1190 1195 1200  
 Ile Ile Ser Ala Ser Met Ala Gly Leu Tyr Thr Ala Gly Lys Ser Leu  
 1205 1210 1215  
 Asn Trp Asp Ala Phe His Lys Glu Asn Glu Ser Leu Arg Val Leu Asn  
 1220 1225 1230  
 Asp Leu Pro Phe Tyr Gly Phe Asp Glu Lys Asn Tyr Trp Leu Gln Tyr  
 1235 1240 1245  
 Thr Gly Asp Trp Leu Leu Tyr Lys Gly Asp Tyr Pro Lys Ala Ile Ala  
 1250 1255 1260  
 Pro Ala Pro Ala Ala Ala Ala Ala Ala Arg Pro Ala Lys Ala Arg Lys  
 1265 1270 1275 1280  
 Tyr Leu Ser Thr Ser Val Gln Gly Ile Val Ser Glu Glu Val Lys Gly  
 1285 1290 1295  
 Lys Thr Val Thr Ile Val Ala Glu Ser Asp Phe Ala His Pro Lys Leu  
 1300 1305 1310  
 Phe Pro Val Ile Ala Gly His Leu Val Asn Gly Ser Gly Leu Cys Pro  
 1315 1320 1325  
 Ser Thr Leu Tyr Ala Asp Met Ala Tyr Thr Leu Gly Gln Leu Gly Val  
 1330 1335 1340  
 Gly Leu Leu Lys Pro Gly Glu Lys Val Asp Ile Asn Ile Gly Thr Met  
 1345 1350 1355 1360  
 Asp Asn Pro Ala Pro Leu Leu Leu Lys Asn Ile Asn Gln Pro Glu Ser  
 1365 1370 1375  
 Gln Ile Val Gln Met Thr Met Lys Ile Asp Leu Asp Ala Arg Lys Ala  
 1380 1385 1390  
 Asp Phe Ala Val Thr Ser Asn Asn Gly Lys Lys Asp Val Thr His Ala  
 1395 1400 1405  
 Lys Cys Val Ile Val Phe Glu Asp Ala Ala Val Trp Lys Glu Gln Trp  
 1410 1415 1420  
 Ser Lys Thr Ser Tyr Leu Ile Gln Ser Arg Ile Asp Met Leu Lys His  
 1425 1430 1435 1440  
 Lys Met Glu Asn Gly Glu Ala Asp Lys Val Ser Arg Ala Met Ala Tyr  
 1445 1450 1455  
 Lys Leu Phe Gly Ala Leu Val Asp Tyr Ala Asp Ile Phe Gln Gly Met  
 1460 1465 1470  
 Gln Asn Val Val Phe Asp Gly Pro Glu Phe Glu Ala Thr Ser Asn Ile  
 1475 1480 1485

## 35

Lys Phe Arg Ala Gly Pro Asn Asp Gly Asp Phe Tyr Phe Ser Pro Tyr  
 1490 1495 1500  
 Phe Ile Asp Ser Ala Cys His Leu Ser Xaa Phe Thr Val Xaa Ala Thr  
 1505 1510 1515 1520  
 Val Xaa Pro Gln Asp Glu Cys Tyr Ile Ser His Gly Trp Ser Ser Leu  
 1525 1530 1535  
 Arg Phe Ile Glu Pro Leu Gln His Asp Gln Gln Tyr Tyr Ala Tyr Leu  
 1540 1545 1550  
 Lys Met Gln Pro Val Ser Gly Ser Lys Xaa Arg Ala Gly Asp Val Tyr  
 1555 1560 1565  
 Val Phe Asn Ala Asp Lys Xaa Val Leu Arg Leu Ala Gly Gly Val Arg  
 1570 1575 1580  
 Phe Gln Cys Ile Pro Arg Lys Leu Met Asp Val Met Met Pro Lys Pro  
 1585 1590 1595 1600  
 Lys Ala Asn Ala Lys Ala Ala Ser Pro Ala Ser Ala Thr Pro Lys Ala  
 1605 1610 1615  
 Ala Pro Thr Val Lys Ala Ser Ser Asn Leu Ala Ala Pro Ala Ser Val  
 1620 1625 1630  
 Glu Leu Pro Thr Pro Lys Val Lys Ala Lys Lys Pro Val Lys Ala Gln  
 1635 1640 1645  
 Lys Ile Lys Ala Pro Ala Pro Pro Lys Gln Ala Ser Ser Gly Ser Leu  
 1650 1655 1660  
 Val Val Arg Ala Phe Glu Ile Ile Ala Lys Glu Ile Asp Val Asp Gln  
 1665 1670 1675 1680  
 Ser Glu Leu Asn Asp Asp Ile Gln Trp Ala Asp Met Gly Val Asp Ser  
 1685 1690 1695  
 Leu Met Ser Leu Thr Ile Ser Gly Lys Phe Arg Glu Asp Leu Asp Leu  
 1700 1705 1710  
 Glu Val Xaa Ser Thr Leu Phe Thr Asp Tyr Ala Ser Val Gly Ala Leu  
 1715 1720 1725  
 Arg Lys His Leu Ser Gly Met Ser Gly Pro Glu Pro Ile Ala Thr Gly  
 1730 1735 1740  
 Asp Ala Ser Ser Val Glu Ser Thr Asp Ser Gly Ser Glu Ser Asp Glu  
 1745 1750 1755 1760  
 Asp Thr Val Glu Ser Gly Ile Thr Thr Pro Asp Thr Glu Xaa Phe Pro  
 1765 1770 1775  
 Ala Lys Xaa Gln Glu Gln Gly Lys Ser Ala Ala Val Glu Ala Met Ala  
 1780 1785 1790

## 36

Gln Pro Pro Ser Ala Glu Gly Gln Asp Met Ile Glu Thr Ile Arg Val  
 1795 1800 1805  
 Val Ile Ala Gln Glu Met Glu Met Asp Leu Ala Glu Ile Thr Asp Xaa  
 1810 1815 1820  
 Thr Asp Leu Ser Asn Leu Gly Met Asp Ser Leu Met Ala Leu Thr Val  
 1825 1830 1835 1840  
 Leu Gly Lys Leu Arg Glu Asp His Asp Ile Asp Leu Asp Pro Thr Ile  
 1845 1850 1855  
 Leu Ala Asp Asn Pro Thr Leu Ala His Leu Arg Lys Ala Leu Gly Leu  
 1860 1865 1870  
 Glu Lys Ala Lys Pro Ala Pro Ala Pro Lys Gln Xaa Val Arg Thr Asn  
 1875 1880 1885  
 Val Val Val Ala Pro Ala Ala Pro Pro Val Xaa Val Val Val Xaa Xaa  
 1890 1895 1900  
 Pro Pro Ala Thr Ser Val Leu Leu Gln Gly Asn Pro Lys Thr Ala Thr  
 1905 1910 1915 1920  
 Xaa Asn Leu Phe Leu Phe Pro Asp Gly Ser Gly Ser Ala Thr Ser Tyr  
 1925 1930 1935  
 Val Ser Ile Pro Ala Ile Asp Ser Xaa Asn Leu Ala Val Tyr Gly Leu  
 1940 1945 1950  
 Asn Cys Pro Phe Met Lys Asp Pro Thr Ser Tyr Thr Cys Gly Ile Xaa  
 1955 1960 1965  
 Ser Val Ser Xaa Leu Tyr Leu Glu Lys Val Leu Xaa Arg Gln Pro Asn  
 1970 1975 1980  
 Gly Pro Tyr Ile Leu Xaa Gly Trp Ser Ala Ser Gly Val Phe Ala Tyr  
 1985 1990 1995 2000  
 Xaa Ile Thr Xaa Gln Leu Xaa Asp Leu Gln Xaa Leu His Pro Asp Lys  
 2005 2010 2015  
 Asn Tyr Thr Val Glu Lys Leu Asn Leu Ile Xaa Ser Pro Cys Pro Ile  
 2020 2025 2030  
 Arg Leu Glu Pro Leu Pro Ala Arg Leu His His Phe Phe Asp Glu Ile  
 2035 2040 2045  
 Gly Leu Leu Gly Thr Gly Thr Gly Lys Thr Pro Asn Trp Leu Leu Pro  
 2050 2055 2060  
 His Phe Glu Tyr Ser Ile Lys Ala Leu Thr Ala Tyr Arg Pro Glu Leu  
 2065 2070 2075 2080  
 Lys Ser Thr Arg Asp Phe Asn Ala Pro Pro Thr Leu Leu Ile Trp Ala  
 2085 2090 2095

## 37

Thr Asp Gly Val Cys Gly Lys Pro Gly Asp Pro Arg Pro Pro Pro Gln  
2100 2105 2110

Ala Asp Asp Pro Lys Ser Met Lys Trp Leu Leu Glu Asn Arg Thr Asp  
2115 2120 2125

Phe Gly Pro Asn Gly Trp Asp Lys Leu Leu Gly Ala Glu Val Cys Lys  
2130 2135 2140

Met Val Thr Val Val Gly Asn His Phe Thr Met Met Lys Pro Pro Val  
2145 2150 2155 2160

Ala Lys Gly Val Gly Gln Tyr Ile Arg Glu Ser Leu Ser Met Xaa Arg  
2165 2170 2175

Ala

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<213> *Aspergillus parasiticus*

<400> 11

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&lt;211&gt; 2109

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<213> *Aspergillus parasiticus*

&lt;400&gt; 12

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Leu Ala Ala Phe Leu Asp Gln Ser His Tyr Val Val Arg Ala Gln Met
          35             40             45
Leu Gln Ser Met Asn Thr Val Asp His Lys Leu Ala Arg Thr Ala Asp
          50             55             60
Leu Arg Gln Met Val Gln Lys Tyr Val Asp Gly Lys Leu Thr Pro Ala
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Phe Arg Thr Ala Leu Val Cys Leu Cys Gln Leu Gly Cys Phe Ile Arg
          85             90             95

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## 40

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 130 135 140  
 Ile Ala Phe Arg Leu Gly Leu Cys Ala Leu Glu Met Arg Asp Arg Val  
 145 150 155 160  
 Asp Gly Cys Ser Asp Asp Arg Gly Asp Pro Trp Ser Thr Ile Val Trp  
 165 170 175  
 Gly Leu Asp Pro Gln Gln Ala Arg Asp Gln Ile Glu Val Phe Cys Arg  
 180 185 190  
 Thr Thr Asn Val Pro Gln Thr Arg Arg Pro Trp Ile Ser Cys Ile Ser  
 195 200 205  
 Lys Asn Ala Ile Thr Leu Ser Gly Ser Pro Ser Thr Leu Arg Ala Phe  
 210 215 220  
 Cys Ala Met Pro Gln Met Ala Gln His Arg Thr Ala Pro Ile Pro Ile  
 225 230 235 240  
 Cys Leu Pro Ala His Asn Gly Ala Leu Phe Thr Gln Ala Asp Ile Thr  
 245 250 255  
 Thr Ile Leu Asp Thr Thr Pro Thr Thr Pro Trp Glu Gln Leu Pro Gly  
 260 265 270  
 Gln Ile Pro Tyr Ile Ser His Val Thr Gly Asn Val Val Gln Thr Ser  
 275 280 285  
 Asn Tyr Arg Asp Leu Ile Glu Val Ala Leu Ser Glu Thr Leu Leu Glu  
 290 295 300  
 Gln Val Arg Leu Asp Leu Val Glu Thr Gly Leu Pro Arg Leu Leu Gln  
 305 310 315 320  
 Ser Arg Gln Val Lys Ser Val Thr Ile Val Pro Phe Leu Thr Arg Met  
 325 330 335  
 Asn Glu Thr Met Ser Asn Ile Leu Pro Asp Ser Phe Ile Ser Thr Glu  
 340 345 350  
 Thr Arg Thr Asp Thr Gly Arg Ala Ile Pro Ala Ser Gly Arg Pro Gly  
 355 360 365  
 Ala Gly Lys Cys Lys Leu Ala Ile Val Ser Met Ser Gly Arg Phe Pro  
 370 375 380  
 Glu Ser Pro Thr Thr Glu Ser Phe Trp Asp Leu Leu Tyr Lys Gly Leu  
 385 390 395 400



## 41

Asp Val Cys Lys Glu Val Pro Arg Arg Arg Trp Asp Ile Asn Thr His  
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 Val Asp Pro Ser Gly Lys Ala Arg Asn Lys Gly Ala Thr Lys Trp Gly  
 420 425 430  
 Cys Trp Leu Asp Phe Ser Gly Asp Phe Asp Pro Arg Phe Phe Gly Ile  
 435 440 445  
 Ser Pro Lys Glu Ala Pro Gln Met Asp Pro Ala Gln Arg Met Ala Leu  
 450 455 460  
 Met Ser Thr Tyr Glu Ala Met Glu Arg Ala Gly Leu Val Pro Asp Thr  
 465 470 475 480  
 Thr Pro Ser Thr Gln Arg Asp Arg Ile Gly Val Phe His Gly Val Thr  
 485 490 495  
 Ser Asn Asp Trp Met Glu Thr Asn Thr Ala Gln Asn Ile Asp Thr Tyr  
 500 505 510  
 Phe Ile Thr Gly Gly Asn Arg Gly Phe Ile Pro Gly Arg Ile Asn Phe  
 515 520 525  
 Cys Phe Glu Phe Ala Gly Pro Ser Tyr Thr Asn Asp Thr Ala Cys Ser  
 530 535 540  
 Ser Ser Leu Ala Ala Ile His Leu Ala Cys Asn Ser Leu Trp Arg Gly  
 545 550 555 560  
 Asp Cys Asp Thr Ala Val Ala Gly Gly Thr Asn Met Ile Tyr Thr Pro  
 565 570 575  
 Asp Gly His Thr Gly Leu Asp Lys Gly Phe Phe Leu Ser Arg Thr Gly  
 580 585 590  
 Asn Cys Lys Pro Tyr Asp Asp Lys Ala Asp Gly Tyr Cys Arg Ala Glu  
 595 600 605  
 Gly Val Gly Thr Val Phe Ile Lys Arg Leu Glu Asp Ala Leu Ala Asp  
 610 615 620  
 Asn Asp Pro Ile Leu Gly Val Ile Leu Asp Ala Lys Thr Asn His Ser  
 625 630 635 640  
 Ala Met Ser Glu Ser Met Thr Arg Pro His Val Gly Ala Gln Ile Asp  
 645 650 655  
 Asn Met Thr Ala Ala Leu Asn Thr Thr Gly Leu His Pro Asn Asp Phe  
 660 665 670  
 Ser Tyr Ile Glu Met His Gly Thr Gly Thr Gln Val Gly Asp Ala Val  
 675 680 685  
 Glu Met Glu Ser Val Leu Ser Val Phe Ala Pro Ser Glu Thr Ala Arg  
 690 695 700

## 42

Lys	Ala	Asp	Gln	Pro	Leu	Phe	Val	Gly	Ser	Ala	Lys	Ala	Asn	Val	Gly	705	710	715	720
His	Gly	Glu	Gly	Val	Ser	Gly	Val	Thr	Ser	Leu	Ile	Lys	Val	Leu	Met	725	730	735	
Met	Met	Gln	His	Asp	Thr	Ile	Pro	Pro	His	Cys	Gly	Ile	Lys	Pro	Gly	740	745	750	
Ser	Lys	Ile	Asn	Arg	Asn	Phe	Pro	Asp	Leu	Gly	Ala	Arg	Asn	Val	His	755	760	765	
Ile	Ala	Phe	Glu	Pro	Lys	Pro	Trp	Pro	Arg	Thr	His	Thr	Pro	Arg	Arg	770	775	780	
Val	Leu	Ile	Asn	Asn	Phe	Ser	Ala	Ala	Gly	Gly	Asn	Thr	Ala	Leu	Ile	785	790	795	800
Val	Glu	Asp	Ala	Pro	Glu	Arg	His	Trp	Pro	Thr	Glu	Lys	Asp	Pro	Arg	805	810	815	
Ser	Ser	His	Ile	Val	Ala	Leu	Ser	Ala	His	Val	Gly	Ala	Ser	Met	Lys	820	825	830	
Thr	Asn	Leu	Glu	Arg	Leu	His	Gln	Tyr	Leu	Leu	Lys	Asn	Pro	His	Thr	835	840	845	
Asp	Leu	Ala	Gln	Leu	Ser	Tyr	Thr	Thr	Thr	Ala	Arg	Arg	Trp	His	Tyr	850	855	860	
Leu	His	Arg	Val	Ser	Val	Thr	Gly	Ala	Ser	Val	Glu	Glu	Val	Thr	Arg	865	870	875	880
Lys	Leu	Glu	Met	Ala	Ile	Gln	Asn	Gly	Asp	Gly	Val	Ser	Arg	Pro	Lys	885	890	895	
Ser	Lys	Pro	Lys	Ile	Leu	Phe	Ala	Phe	Thr	Gly	Gln	Gly	Ser	Gln	Tyr	900	905	910	
Ala	Thr	Met	Gly	Lys	Gln	Val	Tyr	Asp	Ala	Tyr	Pro	Ser	Phe	Arg	Glu	915	920	925	
Asp	Leu	Glu	Lys	Phe	Asp	Arg	Leu	Ala	Gln	Ser	His	Gly	Phe	Pro	Ser	930	935	940	
Phe	Leu	His	Val	Cys	Thr	Ser	Pro	Lys	Gly	Asp	Val	Glu	Glu	Met	Ala	945	950	955	960
Pro	Val	Val	Val	Gln	Leu	Ala	Ile	Thr	Cys	Leu	Gln	Met	Ala	Leu	Thr	965	970	975	
Asn	Leu	Met	Thr	Ser	Phe	Gly	Ile	Arg	Pro	Asp	Val	Thr	Val	Gly	His	980	985	990	
Ser	Leu	Gly	Glu	Phe	Ala	Ala	Leu	Tyr	Ala	Ala	Gly	Val	Leu	Ser	Ala	995	1000	1005	

## 43

Ser Asp Val Val Tyr Leu Val Gly Gln Arg Ala Glu Leu Leu Gln Glu  
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 Arg Cys Gln Arg Gly Thr His Ala Met Leu Ala Val Lys Ala Thr Pro  
 1025 1030 1035 1040  
 Glu Ala Leu Ser Gln Trp Ile Gln Asp His Asp Cys Glu Val Ala Cys  
 1045 1050 1055  
 Ile Asn Gly Pro Glu Asp Thr Val Leu Ser Gly Thr Thr Lys Asn Val  
 1060 1065 1070  
 Ala Glu Val Gln Arg Ala Met Thr Asp Asn Gly Ile Lys Cys Thr Leu  
 1075 1080 1085  
 Leu Lys Leu Pro Phe Ala Phe His Ser Ala Gln Val Gln Pro Ile Leu  
 1090 1095 1100  
 Asp Asp Phe Glu Ala Leu Ala Gln Gly Ala Thr Phe Ala Lys Pro Gln  
 1105 1110 1115 1120  
 Leu Leu Ile Leu Ser Pro Leu Leu Arg Thr Glu Ile His Glu Gln Gly  
 1125 1130 1135  
 Val Val Thr Pro Ser Tyr Val Ala Gln His Cys Arg His Thr Val Asp  
 1140 1145 1150  
 Met Ala Gln Ala Leu Arg Ser Ala Arg Glu Lys Gly Leu Ile Asp Asp  
 1155 1160 1165  
 Lys Thr Leu Val Ile Glu Leu Gly Pro Lys Pro Leu Ile Ser Gly Met  
 1170 1175 1180  
 Val Lys Met Thr Leu Gly Asp Lys Ile Ser Thr Leu Pro Thr Leu Ala  
 1185 1190 1195 1200  
 Pro Asn Lys Ala Ile Trp Pro Ser Leu Gln Lys Ile Leu Thr Ser Val  
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 Tyr Thr Gly Gly Trp Asp Ile Asn Trp Lys Lys Tyr His Ala Pro Phe  
 1220 1225 1230  
 Ala Ser Ser Gln Lys Val Val Asp Leu Pro Ser Tyr Gly Trp Asp Leu  
 1235 1240 1245  
 Lys Asp Tyr Tyr Ile Pro Tyr Gln Gly Asp Trp Cys Leu His Arg His  
 1250 1255 1260  
 Gln Gln Asp Cys Lys Cys Ala Ala Pro Gly His Glu Ile Lys Thr Ala  
 1265 1270 1275 1280  
 Asp Tyr Gln Val Pro Pro Glu Ser Thr Pro His Arg Pro Ser Lys Leu  
 1285 1290 1295  
 Asp Pro Ser Lys Glu Ala Phe Pro Glu Ile Lys Thr Thr Thr Thr Leu  
 1300 1305 1310

## 44

His Arg Val Val Glu Glu Thr Thr Lys Pro Leu Gly Ala Thr Leu Val  
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Val Glu Thr Asp Ile Ser Arg Lys Asp Val Asn Gly Leu Ala Arg Gly  
 1330 1335 1340

His Leu Val Asp Gly Ile Pro Leu Cys Thr Pro Ser Phe Tyr Ala Asp  
 1345 1350 1355 1360

Ile Ala Met Gln Val Gly Gln Tyr Ser Met Gln Arg Leu Arg Ala Gly  
 1365 1370 1375

His Pro Gly Ala Gly Ala Ile Asp Gly Leu Val Asp Val Ser Asp Met  
 1380 1385 1390

Val Val Asp Lys Ala Leu Val Pro His Gly Lys Gly Pro Gln Leu Leu  
 1395 1400 1405

Arg Thr Thr Leu Thr Met Glu Trp Pro Pro Lys Ala Ala Ala Thr Thr  
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Arg Ser Ala Lys Val Lys Phe Ala Thr Tyr Phe Ala Asp Gly Lys Leu  
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Asp Thr Glu His Ala Ser Cys Thr Val Arg Phe Thr Ser Asp Ala Gln  
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Leu Lys Ser Leu Arg Arg Ser Val Ser Glu Tyr Lys Thr His Ile Arg  
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Gln Leu His Asp Gly His Ala Lys Gly Gln Phe Met Arg Tyr Asn Arg  
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Lys Thr Gly Tyr Lys Leu Met Ser Ser Met Ala Arg Phe Asn Pro Asp  
 1490 1495 1500

Tyr Met Leu Leu Asp Tyr Leu Val Leu Asn Glu Ala Glu Asn Glu Ala  
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Ala Ser Gly Val Asp Phe Ser Leu Gly Ser Ser Glu Gly Thr Phe Ala  
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Ala His Pro Ala His Val Asp Ala Ile Thr Gln Val Ala Gly Phe Ala  
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Met Asn Ala Asn Asp Asn Val Asp Ile Glu Lys Gln Val Tyr Val Asn  
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His Gly Trp Asp Ser Phe Gln Ile Tyr Gln Pro Leu Asp Asn Ser Lys  
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Ser Tyr Gln Val Tyr Thr Lys Met Gly Gln Ala Lys Glu Asn Asp Leu  
 1585 1590 1595 1600

Val His Gly Asp Val Val Val Leu Asp Gly Glu Gln Ile Val Ala Phe  
 1605 1610 1615

## 45

Phe Arg Gly Leu Thr Leu Arg Ser Val Pro Arg Gly Ala Leu Arg Val  
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Val Leu Gln Thr Thr Val Lys Lys Ala Asp Arg Gln Leu Gly Phe Lys  
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Thr Met Pro Ser Pro Pro Pro Pro Thr Thr Thr Met Pro Ile Ser Pro  
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Tyr Lys Pro Ala Asn Thr Gln Val Ser Ser Gln Ala Ile Pro Ala Glu  
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Ala Thr His Ser His Thr Pro Pro Gln Pro Lys His Ser Pro Val Pro  
 1685 1690 1695

Glu Thr Ala Gly Ser Ala Pro Ala Ala Lys Gly Val Gly Val Ser Asn  
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Glu Lys Leu Asp Ala Val Met Arg Val Val Ser Glu Glu Ser Gly Ile  
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Ala Leu Glu Glu Leu Thr Asp Asp Ser Asn Phe Ala Asp Met Gly Ile  
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Gly Leu Asp Leu Gly Pro Glu Phe Ser Leu Phe Ile Asp Cys Thr Thr  
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Val Arg Ala Leu Lys Asp Phe Met Leu Gly Ser Gly Asp Ala Gly Ser  
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Gly Ser Asn Val Glu Asp Pro Pro Pro Ser Ala Thr Pro Gly Ile Asn  
 1795 1800 1805

Pro Glu Thr Asp Trp Ser Ser Ser Ala Ser Asp Ser Ile Phe Ala Ser  
 1810 1815 1820

Glu Asp His Gly His Ser Ser Glu Ser Gly Ala Asp Thr Gly Ser Pro  
 1825 1830 1835 1840

Pro Ala Leu Asp Leu Lys Pro Tyr Cys Arg Pro Ser Thr Ser Val Val  
 1845 1850 1855

Leu Gln Gly Leu Pro Met Val Ala Arg Lys Thr Leu Phe Met Leu Pro  
 1860 1865 1870

Asp Gly Gly Gly Ser Ala Phe Ser Tyr Ala Ser Leu Pro Arg Leu Lys  
 1875 1880 1885

Ser Asp Thr Ala Val Val Gly Leu Asn Cys Pro Tyr Ala Arg Asp Pro  
 1890 1895 1900

Glu Asn Met Asn Cys Thr His Gly Ala Met Ile Glu Ser Phe Cys Asn  
 1905 1910 1915 1920

## 46

Glu Ile Arg Arg Arg Gln Pro Arg Gly Pro Tyr His Leu Gly Gly Trp  
                   1925                                  1930                                  1935

Ser Ser Gly Gly Ala Phe Ala Tyr Val Val Ala Glu Ala Leu Val Asn  
                   1940                                  1945                                  1950

Gln Gly Glu Glu Val His Ser Leu Ile Ile Ile Asp Ala Pro Ile Pro  
                   1955                                  1960                                  1965

Gln Ala Met Glu Gln Leu Pro Arg Ala Phe Tyr Glu His Cys Asn Ser  
                   1970                                  1975                                  1980

Ile Gly Leu Phe Ala Thr Gln Pro Gly Ala Ser Pro Asp Gly Ser Thr  
 1985                                  1990                                  1995                                  2000

Glu Pro Pro Ser Tyr Leu Ile Pro His Phe Thr Ala Val Val Asp Val  
                   2005                                  2010                                  2015

Met Leu Asp Tyr Lys Leu Ala Pro Leu His Ala Arg Arg Met Pro Lys  
                   2020                                  2025                                  2030

Val Gly Ile Val Trp Ala Ala Asp Thr Val Met Asp Glu Arg Asp Ala  
                   2035                                  2040                                  2045

Pro Lys Met Lys Gly Met His Phe Met Ile Gln Lys Arg Thr Glu Phe  
                   2050                                  2055                                  2060

Gly Pro Asp Gly Trp Asp Thr Ile Met Pro Gly Ala Ser Phe Asp Ile  
 2065                                  2070                                  2075                                  2080

Val Arg Ala Asp Gly Ala Asn His Phe Thr Leu Met Gln Lys Glu His  
                   2085                                  2090                                  2095

Val Ser Ile Ile Ser Asp Leu Ile Asp Arg Val Met Ala  
                   2100                                  2105

<210> 13  
 <211> 1986  
 <212> PRT  
 <213> *Aspergillus nidulans*

<400> 13

Met Glu Asp Pro Tyr Arg Val Tyr Leu Phe Gly Asp Gln Thr Gly Asp  
   1                                  5                                  10                                  15

Phe Glu Val Gly Leu Arg Arg Leu Leu Gln Ala Lys Asn His Ser Leu  
                   20                                  25                                  30

Leu Ser Ser Phe Leu Gln Arg Ser Tyr His Ala Val Arg Gln Glu Ile  
                   35                                  40                                  45

Ser His Leu Pro Pro Ser Glu Arg Ser Thr Phe Pro Arg Phe Thr Ser  
                   50                                  55                                  60

Ile Gly Asp Leu Leu Ala Arg His Cys Glu Ser Pro Gly Asn Pro Ala  
   65                                  70                                  75                                  80

Ile	Glu	Ser	Val	Leu	Thr	Cys	Ile	Tyr	Gln	Leu	Gly	Cys	Phe	Ile	Asn	85	90	95
Tyr	Tyr	Gly	Asp	Leu	Gly	His	Thr	Phe	Pro	Ser	His	Ser	Gln	Ser	Gln	100	105	110
Leu	Val	Gly	Leu	Cys	Thr	Gly	Leu	Leu	Ser	Cys	Ala	Ala	Val	Ser	Cys	115	120	125
Ala	Ser	Asn	Ile	Gly	Glu	Leu	Leu	Lys	Pro	Ala	Val	Glu	Val	Val	Val	130	135	140
Val	Ala	Leu	Arg	Leu	Gly	Leu	Cys	Val	Tyr	Arg	Val	Arg	Lys	Leu	Phe	145	150	155
Gly	Gln	Asp	Gln	Ala	Ala	Pro	Leu	Ser	Trp	Ser	Ala	Leu	Val	Ser	Gly	165	170	175
Leu	Ser	Glu	Ser	Glu	Gly	Thr	Ser	Leu	Ile	Asp	Lys	Phe	Thr	Arg	Arg	180	185	190
Asn	Val	Ile	Pro	Pro	Ser	Ser	Arg	Pro	Tyr	Ile	Ser	Ala	Val	Cys	Ala	195	200	205
Asn	Thr	Leu	Thr	Ile	Ser	Gly	Pro	Pro	Val	Val	Leu	Asn	Gln	Phe	Leu	210	215	220
Asp	Thr	Phe	Ile	Ser	Gly	Lys	Asn	Lys	Ala	Val	Met	Val	Pro	Ile	His	225	230	235
Gly	Pro	Phe	His	Ala	Ser	His	Leu	Tyr	Glu	Lys	Arg	Asp	Val	Glu	Trp	245	250	255
Ile	Leu	Lys	Ser	Cys	Asn	Val	Glu	Thr	Ile	Arg	Asn	His	Lys	Pro	Arg	260	265	270
Ile	Pro	Val	Leu	Ser	Ser	Asn	Thr	Gly	Glu	Leu	Ile	Val	Val	Glu	Asn	275	280	285
Met	Glu	Gly	Phe	Leu	Lys	Ile	Ala	Leu	Glu	Glu	Ile	Leu	Leu	Arg	Gln	290	295	300
Met	Ser	Trp	Asp	Lys	Val	Thr	Asp	Ser	Cys	Ile	Ser	Ile	Leu	Lys	Ser	305	310	315
Val	Gly	Asp	Asn	Lys	Pro	Lys	Lys	Leu	Leu	Pro	Ile	Ser	Ser	Thr	Ala	325	330	335
Thr	Gln	Ser	Leu	Phe	Asn	Ser	Leu	Lys	Lys	Ser	Asn	Leu	Val	Asn	Ile	340	345	350
Glu	Val	Asp	Gly	Gly	Ile	Ser	Asp	Phe	Ala	Ala	Glu	Thr	Gln	Leu	Val	355	360	365
Asn	Gln	Thr	Gly	Arg	Ala	Glu	Leu	Ser	Lys	Ile	Ala	Ile	Ile	Gly	Met	370	375	380

48

Ser Gly Arg Phe Pro Glu Ala Asp Ser Pro Gln Asp Phe Trp Asn Leu		
385	390	395 400
Leu Tyr Lys Gly Leu Asp Val His Arg Lys Val Pro Glu Asp Arg Trp		
	405	410 415
Asp Ala Asp Ala His Val Asp Leu Thr Gly Thr Ala Thr Asn Thr Ser		
	420	425 430
Lys Val Pro Tyr Gly Cys Trp Ile Arg Glu Pro Gly Leu Phe Asp Pro		
	435	440 445
Arg Phe Phe Asn Met Ser Pro Arg Glu Ala Leu Gln Ala Asp Pro Ala		
	450	455 460
Gln Arg Leu Ala Leu Leu Thr Ala Tyr Glu Ala Leu Glu Gly Ala Gly		
	465	470 475 480
Phe Val Pro Asp Ser Thr Pro Ser Thr Gln Arg Asp Arg Val Gly Ile		
	485	490 495
Phe Tyr Gly Met Thr Ser Asp Asp Tyr Arg Glu Val Asn Ser Gly Gln		
	500	505 510
Asp Ile Asp Thr Tyr Phe Ile Pro Gly Gly Asn Arg Ala Phe Thr Pro		
	515	520 525
Gly Arg Ile Asn Tyr Tyr Phe Lys Phe Ser Gly Pro Ser Val Ser Val		
	530	535 540
Asp Thr Ala Cys Ser Ser Ser Leu Ala Ala Ile His Leu Ala Cys Asn		
	545	550 555 560
Ser Ile Trp Arg Asn Asp Cys Asp Thr Ala Ile Thr Gly Gly Val Asn		
	565	570 575
Ile Leu Thr Asn Pro Asp Asn His Ala Gly Leu Asp Arg Gly His Phe		
	580	585 590
Leu Ser Arg Thr Gly Asn Cys Asn Thr Phe Asp Asp Gly Ala Asp Gly		
	595	600 605
Tyr Cys Arg Ala Asp Gly Val Gly Thr Val Val Leu Lys Arg Leu Glu		
	610	615 620
Asp Ala Leu Ala Asp Asn Asp Pro Ile Leu Gly Val Ile Asn Gly Ala		
	625	630 635 640
Tyr Thr Asn His Ser Ala Glu Ala Val Ser Ile Thr Arg Pro His Val		
	645	650 655
Gly Ala Gln Ala Phe Ile Phe Lys Lys Leu Leu Asn Glu Ala Asn Val		
	660	665 670
Asp Pro Lys Asn Ile Ser Tyr Ile Glu Met His Gly Thr Gly Thr Gln		
	675	680 685



Ala Gly Asp Ala Val Glu Met Gln Ser Val Leu Asp Val Phe Ala Pro  
 690 695 700  
 Asp His Arg Arg Gly Pro Gly Gln Ser Leu His Leu Gly Ser Ala Lys  
 705 710 715 720  
 Ser Asn Ile Gly His Gly Glu Ser Ala Ser Gly Val Thr Ser Leu Val  
 725 730 735  
 Lys Val Leu Leu Met Met Lys Glu Asn Met Ile Pro Pro His Cys Gly  
 740 745 750  
 Ile Lys Thr Lys Ile Asn His Asn Phe Pro Thr Asp Leu Ala Gln Arg  
 755 760 765  
 Asn Val His Ile Ala Leu Gln Pro Thr Ala Trp Asn Arg Pro Ser Phe  
 770 775 780  
 Gly Lys Arg Gln Ile Phe Leu Asn Asn Phe Ser Ala Ala Gly Gly Asn  
 785 790 795 800  
 Thr Ala Leu Leu Leu Glu Asp Gly Pro Val Ser Asp Pro Glu Gly Glu  
 805 810 815  
 Asp Lys Arg Arg Thr His Val Ile Thr Leu Ser Ala Arg Ser Gln Thr  
 820 825 830  
 Ala Leu Gln Asn Asn Ile Asp Ala Leu Cys Gln Tyr Ile Ser Glu Gln  
 835 840 845  
 Glu Lys Thr Phe Gly Val Lys Asp Ser Asn Ala Leu Pro Ser Leu Ala  
 850 855 860  
 Tyr Thr Thr Thr Ala Arg Arg Ile His His Pro Phe Arg Val Thr Ala  
 865 870 875 880  
 Ile Gly Ser Ser Phe Gln Glu Met Arg Asp Ser Leu Ile Ala Ser Ser  
 885 890 895  
 Arg Lys Glu Phe Val Ala Val Pro Ala Lys Thr Pro Gly Ile Gly Phe  
 900 905 910  
 Leu Phe Thr Gly Gln Gly Ala Gln Tyr Ala Ala Met Gly Lys Gln Leu  
 915 920 925  
 Tyr Glu Asp Cys Ser His Phe Arg Ser Ala Ile Glu His Leu Asp Cys  
 930 935 940  
 Ile Ser Gln Gly Gln Asp Leu Pro Ser Ile Leu Pro Leu Val Asp Gly  
 945 950 955 960  
 Ser Leu Pro Leu Ser Glu Leu Ser Pro Val Val Val Gln Leu Gly Thr  
 965 970 975  
 Thr Cys Val Gln Met Ala Leu Ser Ser Phe Trp Ala Ser Leu Gly Ile  
 980 985 990

## 50

Thr Pro Ser Phe Val Leu Gly His Ser Leu Gly Asp Phe Ala Ala Met  
 995 1000 1005  
 Asn Ala Ala Gly Val Leu Ser Thr Ser Asp Thr Ile Tyr Ala Cys Gly  
 1010 1015 1020  
 Arg Arg Ala Gln Leu Leu Thr Glu Arg Cys Gln Pro Gly Thr His Ala  
 1025 1030 1035 1040  
 Met Leu Ala Ile Lys Ala Pro Leu Val Glu Val Lys Gln Leu Leu Asn  
 1045 1050 1055  
 Glu Lys Val His Asp Met Ala Cys Ile Asn Ser Pro Ser Glu Thr Val  
 1060 1065 1070  
 Ile Ser Gly Pro Lys Ser Ser Ile Asp Glu Leu Ser Arg Ala Cys Ser  
 1075 1080 1085  
 Glu Lys Gly Leu Lys Ser Thr Ile Leu Thr Val Pro Tyr Ala Phe His  
 1090 1095 1100  
 Ser Ala Gln Val Glu Pro Ile Leu Glu Asp Leu Glu Lys Ala Leu Gln  
 1105 1110 1115 1120  
 Gly Ile Thr Phe Asn Lys Pro Ser Val Pro Phe Val Ser Ala Leu Leu  
 1125 1130 1135  
 Gly Glu Val Ile Thr Glu Ala Gly Ser Asn Ile Leu Asn Ala Glu Tyr  
 1140 1145 1150  
 Leu Val Arg His Cys Arg Glu Thr Val Asn Phe Leu Ser Ala Phe Glu  
 1155 1160 1165  
 Ala Val Arg Asn Ala Lys Leu Gly Gly Asp Gln Thr Leu Trp Leu Glu  
 1170 1175 1180  
 Val Gly Pro His Thr Val Cys Ser Gly Met Val Lys Ala Thr Leu Gly  
 1185 1190 1195 1200  
 Pro Gln Thr Thr Thr Met Ala Ser Leu Arg Arg Asp Glu Asp Thr Trp  
 1205 1210 1215  
 Lys Val Leu Ser Asn Ser Leu Ser Ser Leu Tyr Leu Ala Gly Val Asp  
 1220 1225 1230  
 Ile Asn Trp Lys Gln Tyr His Gln Asp Phe Ser Ser Ser His Arg Val  
 1235 1240 1245  
 Leu Pro Leu Pro Thr Tyr Lys Trp Asp Leu Lys Asn Tyr Trp Ile Pro  
 1250 1255 1260  
 Tyr Arg Asn Asn Phe Cys Leu Thr Lys Gly Ser Ser Met Ser Ala Ala  
 1265 1270 1275 1280  
 Ser Ala Ser Leu Gln Pro Thr Phe Leu Thr Thr Ser Ala Gln Arg Val  
 1285 1290 1295

## 51

Val Glu Ser Arg Asp Asp Gly Leu Thr Ala Thr Val Val Val His Asn  
 1300 1305 1310  
 Asp Ile Ala Asp Pro Asp Leu Asn Arg Val Ile Gln Gly His Lys Val  
 1315 1320 1325  
 Asn Gly Ala Ala Leu Cys Pro Ser Ser Leu Tyr Ala Asp Ser Ala Gln  
 1330 1335 1340  
 Thr Leu Ala Glu Tyr Leu Ile Glu Lys Tyr Lys Pro Glu Leu Lys Gly  
 1345 1350 1355 1360  
 Ser Gly Leu Asp Val Cys Asn Val Thr Val Pro Lys Pro Leu Ile Ala  
 1365 1370 1375  
 Lys Thr Gly Lys Glu Gln Phe Arg Ile Ser Ala Thr Ala Asn Trp Val  
 1380 1385 1390  
 Asp Lys His Val Ser Val Gln Val Phe Ser Val Thr Ala Glu Gly Lys  
 1395 1400 1405  
 Lys Leu Ile Asp His Ala His Cys Glu Val Lys Leu Phe Asp Cys Met  
 1410 1415 1420  
 Ala Ala Asp Leu Glu Trp Lys Arg Gly Ser Tyr Leu Val Lys Arg Ser  
 1425 1430 1435 1440  
 Ile Glu Leu Leu Glu Asn Ser Ala Val Lys Gly Asp Ala His Arg Leu  
 1445 1450 1455  
 Arg Arg Gly Met Val Tyr Lys Leu Phe Ser Ala Leu Val Asp Tyr Asp  
 1460 1465 1470  
 Glu Asn Tyr Gln Ser Ile Arg Glu Val Ile Leu Asp Ser Glu His His  
 1475 1480 1485  
 Glu Ala Thr Ala Leu Val Lys Phe Gln Ala Pro Gln Ala Asn Phe His  
 1490 1495 1500  
 Arg Asn Pro Tyr Trp Ile Asp Ser Phe Gly His Leu Ser Gly Phe Ile  
 1505 1510 1515 1520  
 Met Asn Ala Ser Asp Gly Thr Asp Ser Lys Ser Gln Val Phe Val Asn  
 1525 1530 1535  
 His Gly Trp Asp Ser Met Arg Cys Leu Lys Lys Phe Ser Ala Asp Val  
 1540 1545 1550  
 Thr Tyr Arg Thr Tyr Val Arg Met Gln Pro Trp Arg Asp Ser Ile Trp  
 1555 1560 1565  
 Ala Gly Asn Val Tyr Ile Phe Glu Gly Asp Asp Ile Ile Ala Val Phe  
 1570 1575 1580  
 Gly Gly Val Lys Phe Gln Ala Leu Ser Arg Lys Ile Leu Asp Ile Ala  
 1585 1590 1595 1600

## 52

Leu Pro Pro Ala Gly Leu Ser Lys Ala Gln Thr Ser Pro Ile Gln Ser  
 1605 1610 1615  
 Ser Ala Pro Gln Lys Pro Ile Glu Thr Ala Lys Pro Thr Ser Arg Pro  
 1620 1625 1630  
 Ala Pro Pro Val Thr Met Lys Ser Phe Val Lys Lys Ser Ala Gly Pro  
 1635 1640 1645  
 Ser Val Val Val Arg Ala Leu Asn Ile Leu Ala Ser Glu Val Gly Leu  
 1650 1655 1660  
 Ser Glu Ser Asp Met Ser Asp Asp Leu Val Phe Ala Asp Tyr Gly Val  
 1665 1670 1675 1680  
 Asp Ser Leu Leu Ser Leu Thr Val Thr Gly Lys Tyr Arg Glu Glu Leu  
 1685 1690 1695  
 Asn Leu Asp Met Asp Ser Ser Val Phe Ile Glu His Pro Thr Val Gly  
 1700 1705 1710  
 Asp Phe Lys Arg Phe Val Thr Gln Leu Ser Pro Ser Val Ala Ser Asp  
 1715 1720 1725  
 Ser Ser Ser Thr Asp Arg Glu Ser Glu Tyr Ser Phe Asn Gly Asp Ser  
 1730 1735 1740  
 Cys Ser Gly Leu Ser Ser Pro Ala Ser Pro Gly Thr Val Ser Pro Pro  
 1745 1750 1755 1760  
 Asn Glu Lys Val Ile Gln Ile His Glu Asn Gly Thr Met Lys Glu Ile  
 1765 1770 1775  
 Arg Ala Ile Ile Ala Asp Glu Ile Gly Val Ser Ala Asp Glu Ile Lys  
 1780 1785 1790  
 Ser Asp Glu Asn Leu Asn Glu Leu Gly Met Asp Ser Leu Leu Ser Leu  
 1795 1800 1805  
 Thr Val Leu Gly Lys Ile Arg Glu Ser Leu Asp Met Asp Leu Pro Gly  
 1810 1815 1820  
 Glu Phe Phe Ile Glu Asn Gln Thr Leu Asp Gln Ile Glu Thr Ala Leu  
 1825 1830 1835 1840  
 Asp Leu Lys Pro Lys Ala Val Pro Thr Ala Val Pro Gln Ser Gln Pro  
 1845 1850 1855  
 Ile Thr Leu Pro Gln Ser Gln Ser Thr Lys Gln Leu Ser Thr Arg Pro  
 1860 1865 1870  
 Thr Ser Ser Ser Asp Asn His Pro Pro Ala Thr Ser Ile Leu Leu Gln  
 1875 1880 1885  
 Gly Asn Pro Arg Thr Ala Ser Lys Thr Leu Phe Leu Phe Pro Asp Gly  
 1890 1895 1900

## 53

Ser Gly Ser Ala Thr Ser Tyr Ala Thr Ile Pro Gly Val Ser Pro Asn  
1905 1910 1915 1920

Val Ala Val Tyr Gly Leu Asn Cys Pro Tyr Met Lys Ala Pro Glu Lys  
1925 1930 1935

Leu Thr Cys Ser Leu Asp Ser Leu Thr Thr Pro Tyr Leu Ala Glu Ile  
1940 1945 1950

Arg Arg Arg Gln Pro Thr Gly Pro Tyr Asn Leu Gly Gly Trp Ser Gln  
1955 1960 1965

Ala Gly Ser Ala His Thr Thr Arg His Ala Ser Ser Tyr Cys Ser Arg  
1970 1975 1980

Ala Lys  
1985

<210> 14  
<211> 53  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: Primer

<400> 14  
atgaagcttg gggtttgagg gccaatggaa cgaaactagt gtaccacttg acc 53

<210> 15  
<211> 28  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: Primer

<400> 15  
gacagatctg gcgccattcg ccattcag 28

<210> 16  
<211> 20  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: Primer

<400> 16  
ggaatcggtc aatacactac 20

<210> 17  
<211> 33  
<212> DNA  
<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 17

tgtagatctc tattcctttg ccctcggacg agt

33

<210> 18

<211> 35

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 18

ggccgccacg gatattcttg ccaaagaatt cctgg

35

<210> 19

<211> 35

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 19

cgggtgcctat agaaccggtt tcttaaggac cgcg

35

<210> 20

<211> 19

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 20

gayccmgtty ttyaayatg

19

<210> 21

<211> 17

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 21

gtccgtccrt gcatytc

17

<210> 22

<211> 34

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 22

ataagaatgc ggccgcaatg gccctcgaaa cagc

34

<210> 23

<211> 29

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 23

aaatggcgcg ccgcgcccag aatgacacc

29

<210> 24

<211> 23

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 24

tgccacctgt agtctgcaat cag

23

<210> 25

<211> 24

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 25

tgactaaccc tgacaacttc gctg

24

<210> 26

<211> 19

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 26

ccaggatccg actgctcag

19

<210> 27

<211> 21

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 27

ctacatcgag atgcacggca c

21

<210> 28

<211> 16

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 28

ngtcgaswga nawgaa

16

<210> 29

<211> 16

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 29

gtncgaswca nawgtt

16

<210> 30

<211> 16

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 30

wgtgnagwan canaga

16

<210> 31

<211> 15

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 31

ntcgastwts gwggtt

15

<210> 32

<211> 16

<212> DNA

<213> Artificial Sequence



<220>

<223> Description of Artificial Sequence: Primer

<400> 32

tgwgnagwan casaga

16

<210> 33

<211> 16

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 33

agwgnagwan cawagg

16

<210> 34

<211> 14

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 34

cawcgngaa sgaa

14

<210> 35

<211> 14

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 35

tcstcgnact wgga

14

<210> 36

<211> 23

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 36

ttgttactgg agaggtaatg aag

23

<210> 37

<211> 23

<212> DNA

<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: Primer

<400> 37  
tgagacagat ctgcgagcc ctc 23

<210> 38  
<211> 22  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: Primer

<400> 38  
atgtctccaa aggaagctga gc 22

<210> 39  
<211> 22  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: Primer

<400> 39  
tcgagtgatg gatactgctt cg 22

<210> 40  
<211> 29  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: Primer

<400> 40  
cggctacact agaaggacag tatttggtta 29

<210> 41  
<211> 30  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: Primer

<400> 41  
gtcaggcaac tatggatgaa cgaaatagac 30

<210> 42  
<211> 25  
<212> DNA  
<213> Artificial Sequence

59

&lt;220&gt;

&lt;223&gt; Description of Artificial Sequence: Primer

&lt;400&gt; 42

acccatctca taaataacgt catgc

25

&lt;210&gt; 43

&lt;211&gt; 23

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

&lt;223&gt; Description of Artificial Sequence: Primer

&lt;400&gt; 43

caactctatc agagcttggt tga

23

&lt;210&gt; 44

&lt;211&gt; 30

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

&lt;223&gt; Description of Artificial Sequence: Primer

&lt;400&gt; 44

cccgaattca tgagctttgt tcaaataagg

30

&lt;210&gt; 45

&lt;211&gt; 39

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

&lt;223&gt; Description of Artificial Sequence: Primer

&lt;400&gt; 45

ttattctaga ttttccatgg gaatggatac agtcttacg

39

&lt;210&gt; 46

&lt;211&gt; 33

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

&lt;223&gt; Description of Artificial Sequence: Primer

&lt;400&gt; 46

cgccaccatg gtgagcaagg gcgaggagct gtt

33

&lt;210&gt; 47

&lt;211&gt; 39

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

&lt;223&gt; Description of Artificial Sequence: Primer

&lt;400&gt; 47

tatgatctag agtcgcgggc gctttacttg tacagctcg

39

&lt;210&gt; 48

&lt;211&gt; 25

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

&lt;223&gt; Description of Artificial Sequence: Primer

&lt;400&gt; 48

gcttctaatac cgtactagtg gatca

25

&lt;210&gt; 49

&lt;211&gt; 25

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

&lt;223&gt; Description of Artificial Sequence: Primer

&lt;400&gt; 49

ctttgatctt ttctacgggg tctga

25